

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

**PATIENT**

**DISEASE** Lung adenocarcinoma  
**NAME** Not Given  
**DATE OF BIRTH** Not Given  
**SEX** Not Given  
**MEDICAL RECORD #** Not Given

**PHYSICIAN**

**ORDERING PHYSICIAN** Not Given  
**MEDICAL FACILITY** Not Given  
**ADDITIONAL RECIPIENT** Not Given  
**MEDICAL FACILITY ID** Not Given  
**PATHOLOGIST** Not Given

**SPECIMEN**

**SPECIMEN SITE** Not Given  
**SPECIMEN ID** Not Given  
**SPECIMEN TYPE** Not Given  
**DATE OF COLLECTION** Not Given  
**SPECIMEN RECEIVED** Not Given

**Biomarker Findings**
**Microsatellite status - MS-Stable**
**Tumor Mutational Burden - TMB-Low (4 Muts/Mb)**
**Genomic Findings**

*For a complete list of the genes assayed, please refer to the Appendix.*

**ALK EML4-ALK fusion (Variant 1)**
**CCND1 amplification**
**FGF19 amplification**
**FGF3 amplification**
**FGF4 amplification**
**NFKBIA amplification**
**NKX2-1 amplification**
**TP53 R306\***

**7 Disease-relevant genes with no reportable alterations: EGFR, KRAS, BRAF, MET, RET, ERBB2, ROS1**

**7 Therapies with Clinical Benefit**
**14 Clinical Trials**
**0 Therapies with Lack of Response**
**BIOMARKER FINDINGS**
**Microsatellite status - MS-Stable**
**Tumor Mutational Burden - TMB-Low (4 Muts/Mb)**
**GENOMIC FINDINGS**
**ALK - EML4-ALK fusion (Variant 1)**
**10 Trials** see p. 11

**CCND1 - amplification**
**4 Trials** see p. 14

**ACTIONABILITY**
**No therapies or clinical trials.** see Biomarker Findings section

**No therapies or clinical trials.** see Biomarker Findings section

**THERAPIES WITH CLINICAL BENEFIT (IN PATIENT'S TUMOR TYPE)**

Alectinib

Brigatinib

Ceritinib

Crizotinib

None

**THERAPIES WITH CLINICAL BENEFIT (IN OTHER TUMOR TYPE)**

None

Abemaciclib

Palbociclib

Ribociclib

**GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS**

*For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.*

<b>FGF19</b> - amplification .....	p. 5	<b>NFKBIA</b> - amplification .....	p. 6
<b>FGF3</b> - amplification .....	p. 5	<b>NKX2-1</b> - amplification .....	p. 6
<b>FGF4</b> - amplification .....	p. 6	<b>TP53</b> - R306* .....	p. 7

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

QRF# XXXXXXXX

**BIOMARKER FINDINGS**
**BIOMARKER**

## Microsatellite status

**CATEGORY**

MS-Stable

**POTENTIAL TREATMENT STRATEGIES**

On the basis of clinical evidence, microsatellite stable (MSS) tumors are significantly less likely than MSI-high (MSI-H) tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>1-3</sup>, including approved therapies nivolumab and pembrolizumab<sup>4-5</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,

$p=0.001$ )<sup>6</sup>. Pembrolizumab therapy resulted in a significantly lower objective response rate (ORR) in MSS colorectal cancer (CRC) compared with MSI-H CRC (0% vs. 40%)<sup>5</sup>. Similarly, a clinical study of nivolumab, alone or in combination with ipilimumab, in patients with CRC reported a significantly higher response rate in patients with MSI-H tumors than those without<sup>4</sup>.

**FREQUENCY & PROGNOSIS**

MSI-high (MSI-H) has been reported at various frequencies in non-small cell lung cancer (NSCLC) as well as in small cell lung cancer<sup>7-12</sup>. One study observed MSI-H in 0.8% (4/480) of lung adenocarcinoma cases; the MSI-H tumors occurred in patients with smoking history, and 3 of the 4 MSI-H cases had nonsynchronous carcinomas in other organs, although none of the patients were diagnosed with Lynch syndrome<sup>7</sup>.

**FINDING SUMMARY**

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>13</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>13-15</sup>. The tumor seen here is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>16-18</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins<sup>13,15,17-18</sup>.

QRF# XXXXXXXX

**BIOMARKER FINDINGS**
**BIOMARKER**

# Tumor Mutational Burden

**CATEGORY**
**TMB-Low (4 Muts/Mb)**
**POTENTIAL TREATMENT STRATEGIES**

On the basis of emerging clinical evidence, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-CTLA-4<sup>19</sup>, anti-PD-L1<sup>20-23</sup>, and anti-PD-1 therapies<sup>5,24-25</sup>; FDA-approved agents include ipilimumab, atezolizumab, avelumab, durvalumab, pembrolizumab, and nivolumab. In multiple solid tumor types, higher mutational burden has corresponded with response and improved prognosis. Pembrolizumab improved progression-free survival (14.5 vs. 3.4-3.7 months) for patients with non-small cell lung cancer (NSCLC) and higher mutational load (greater than 200 nonsynonymous mutations; hazard ratio = 0.19)<sup>25</sup>. In studies of patients with either NSCLC or colorectal cancer (CRC), patients whose tumors harbored elevated mutational burden reported higher overall response rates to pembrolizumab<sup>5,24-25</sup>. Anti-PD-1 therapies have achieved clinical benefit for certain patients with high mutational burden, including 3 patients with endometrial adenocarcinoma who reported sustained partial responses (PRs) following treatment with pembrolizumab<sup>26</sup> or nivolumab<sup>27</sup>, a patient with hypermutant glioblastoma who obtained clinical benefit from pembrolizumab<sup>28</sup>, 2 pediatric patients with

biallelic mismatch repair deficiency-associated ultrahypermutant glioblastoma who experienced clinically and radiologically significant responses to nivolumab<sup>29</sup>, and 2 patients with microsatellite-stable rectal cancers, 1 who achieved an ongoing PR to pembrolizumab and the other an ongoing complete response to nivolumab<sup>30</sup>. For patients with melanoma, mutational load was associated with long-term clinical benefit from ipilimumab<sup>19,31</sup> and anti-PD-1/anti-PD-L1 treatments<sup>21</sup>. For patients with metastatic urothelial carcinoma, those who responded to atezolizumab treatment had a significantly increased mutational load (12.4 mutations [mut] per megabase [Mb]) compared to nonresponders (6.4 muts/Mb)<sup>20</sup>, and mutational load of 16 muts/Mb or higher was associated with significantly longer overall survival<sup>22</sup>. In a retrospective analysis of 17 solid tumor types (comprised of 47% NSCLC, 40% mUC, and 13% encompassing 15 other solid tumors), a TMB of  $\geq 16$  muts/Mb associated with an objective response rate to atezolizumab of 30% vs. 14% for chemotherapy alone<sup>32</sup>.

**FREQUENCY & PROGNOSIS**

Intermediate TMB has been reported in 30-31% of non-small cell lung carcinomas (NSCLC), including 30% of adenocarcinomas and 41% of squamous cell carcinomas (SCC) (Spigel et al., 2016; ASCO Abstract 9017). Intermediate TMB was frequently observed in NSCLC with BRAF (31%) or KRAS (39%) mutation (Spigel et al., 2016; ASCO Abstract 9017). Although some studies have reported a lack of association between smoking and mutational burden in NSCLC (Schwartz et al., 2016; ASCO Abstract 8533)<sup>66,67</sup>, several other large studies did find a strong association with increased

Low TMB is observed more commonly in non-small cell lung carcinomas (NSCLC) harboring known driver mutations (EGFR, ALK, ROS1, or MET) with the exception of BRAF or KRAS mutations, which are observed in approximately half of intermediate-high TMB cases<sup>33</sup>. Although some studies have reported a lack of association between smoking and mutational burden in NSCLC<sup>34-36</sup>, several other large studies did find a strong association with increased TMB<sup>37-40</sup>. A large study of Chinese patients with lung adenocarcinoma reported a shorter median overall survival (OS) for tumors with a higher number of mutations in a limited gene set compared with lower mutation number (48.4 vs. 61.0 months)<sup>35</sup>.

**FINDING SUMMARY**

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>41-42</sup> and cigarette smoke in lung cancer<sup>25,43</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>44-48</sup>, and microsatellite instability (MSI)<sup>44,47-48</sup>. The tumor seen here harbors a low TMB. Compared to patients with tumors harboring higher TMB levels, patients with tumors harboring low TMB levels have experienced lower rates of clinical benefit from treatment with immune checkpoint inhibitors, including anti-CTLA-4 therapy in melanoma<sup>19</sup>, anti-PD-L1 therapy in urothelial carcinoma<sup>20</sup>, and anti-PD-1 therapy in non-small cell lung cancer and colorectal cancers<sup>5,25</sup>.

QRF# XXXXXXXX

## GENOMIC FINDINGS

GENE  
**ALK**

ALTERATION  
**EML4-ALK fusion (Variant 1)**

## POTENTIAL TREATMENT STRATEGIES

The ALK inhibitors crizotinib, ceritinib, brigatinib, and alectinib have shown significant clinical activity for patients with non-small cell lung cancer (NSCLC) whose tumors test positive for ALK rearrangement<sup>49-50, 51-56</sup>. As first-line treatment, crizotinib improved overall survival (OS) relative to chemotherapy (HR=0.35) for patients with ALK+ advanced NSCLC<sup>57</sup>. Crizotinib has also shown activity in ALKmutant neuroblastoma<sup>58-59</sup>. Preclinically, ALK activating point mutations are crizotinib-sensitive<sup>60-61</sup>. A Phase 1 study of ceritinib in ALK-rearranged NSCLC reported overall survival (OS) of 72% (60/83) for patients who were ALK inhibitor-naïve and median progression-free survival (PFS) of 18.4 months, versus an OS of 56% (92/163) and PFS of 6.9 months for those who were previously treated<sup>62</sup>. A Phase 1/2 study of brigatinib for patients with ALK-rearranged NSCLC reported confirmed ORRs of 62% (44/71) and 100% (8/8) for crizotinib-treated and crizotinib-naïve patients, respectively<sup>53</sup>. Antitumor activity was also seen in the central nervous system (CNS), a common site of failure during crizotinib treatment<sup>53,63-64</sup>. Alectinib combined with atezolizumab led to an ORR of 81% (17/21) as first-line treatment for PD-L1 unselected, ALK+ NSCLC<sup>65</sup>. Lorlatinib led to an ORR of 73% (43/59), 39% (11/28), and 39% (43/111), and intracranial ORR of 68% (25/37), 46% (6/13), and 47% (38/81), for patients with NSCLC previously treated with crizotinib, one

prior ALK inhibitor, or 2-3 prior ALK inhibitors, respectively<sup>66</sup>. For patients whose tumors harbored one or more ALK kinase domain mutations, lorlatinib led to responses for 64% (29/45), including 58% (11/19) for those with the ALK G1202R resistance mutation<sup>67</sup>; G1202 therefore does not appear to represent a major mechanism of lorlatinib resistance<sup>68-69</sup>. Lorlatinib led to complete resolution of intrathecal metastases and stabilization of CNS metastases for a heavily pretreated patient with ALK+ NSCLC<sup>70</sup>, and its use in the fourth-line setting led to disappearance of leptomeningeal disease for a patient with ALK-rearranged metastatic inflammatory myofibroblastic sarcoma<sup>71</sup>. The combination of lorlatinib and the PD-L1 inhibitor avelumab led to a confirmed response rate of 46.4% [12 partial responses (PRs), 1 complete response] for the 28 patients with ALK+ NSCLC who were treated<sup>72</sup>. Ensartinib treatment for ALK+ NSCLC led to ORRs of 80%, 69%, and 64% for patients who were treatment-naïve, crizotinib refractory, or for intracranial metastases, respectively<sup>73</sup>. Phase 1 studies of the ALK/ROS1/TRK inhibitor entrectinib have reported responses for 4/7 (57%) kinase inhibitor-naïve patients with ALK-rearranged solid tumors, including patients with NSCLC, renal cell carcinoma, and colorectal cancer; as well as for 1 patient with ALK F1245V mutant neuroblastoma but in 0/13 patients with ALK fusion-positive tumors previously treated with an ALK inhibitor and in none of the other patients with ALK non-fusion alterations<sup>74</sup>. A Phase 2 trial of the HSP90 inhibitor ganetespib reported PRs in a small number of patients with ALK-rearranged NSCLC<sup>75</sup>.

## FREQUENCY &amp; PROGNOSIS

The EML4-ALK gene fusion has been observed in approximately 3-7% of non-small cell lung cancer (NSCLC) cases, more frequently in younger patients, non-smokers, males, and

patients of Asian heritage<sup>76-82</sup>. Other rearrangements involving ALK have also been described in lung cancer<sup>83-84</sup>. EML4-ALK fusions have been reported to be a significant indicator of poor prognosis in advanced stage NSCLC<sup>82</sup>.

## FINDING SUMMARY

ALK encodes a receptor tyrosine kinase, a member of the insulin receptor superfamily, whose activation induces the downstream pathways associated with cell survival, angiogenesis, and cell proliferation<sup>85</sup>. Different EML4-ALK variants have been identified in cancer, all of which contain the intracellular tyrosine kinase domain of ALK<sup>86</sup>. The most commonly observed rearrangements consist of ALK exon 20 fused to a variety of breakpoints in EML4: exon 13 (variant 1, 33-54% of cases)<sup>87-89</sup>, exon 20 (variant 2, 10-12% of cases)<sup>87-89</sup>, exon 6 (variant 3 a/b, 26-30% of cases)<sup>52,87-88,90</sup>, exon 15 (variant 4, 2% of cases)<sup>76,91-92</sup>, exon 18 (variant 5, 1.6-3% of cases)<sup>89,91</sup>, exon 2 (variant 5 a/b, 1-2% of cases)<sup>87,92-94</sup>, and exon 17 (variant 8 a/b, <1%)<sup>89,91,95</sup>. All of these variants have been characterized as, or are predicted to be, activating and sensitive to ALK inhibitors, including crizotinib and ceritinib<sup>88,90,96</sup>; however, variants 3a/b are less sensitive to crizotinib in vitro<sup>88</sup>. Although EML4-ALK variant 1 was associated with significantly longer median progression-free survival (11 months vs. 4.2 months) in a small study of crizotinib-treated non-small cell lung cancer (NSCLC)<sup>97</sup>, other studies have not found a correlation between EML4-ALK variants and response to crizotinib in NSCLC<sup>52,89</sup>.

QRF# XXXXXXXX

**GENOMIC FINDINGS**
**GENE**  
**CCND1**
**ALTERATION**  
 amplification

**POTENTIAL TREATMENT STRATEGIES**

Amplification or overexpression of CCND1 may predict sensitivity to CDK4/6 inhibitors, such as FDA-approved abemaciclib, palbociclib,

and ribociclib<sup>98-99, 100-101, 102-105</sup>. Clinical benefit has been reported for patients with solid tumors with CCND1 amplification or expression in response to treatment with palbociclib<sup>106</sup>, ribociclib<sup>98-99, 100, 104</sup>, and abemaciclib<sup>105</sup>.

**FREQUENCY & PROGNOSIS**

In the TCGA dataset, amplification of CCND1 has been found in 4.3% of lung adenocarcinoma cases<sup>107</sup>. Other studies have reported CCND1 amplification in 3-25% of lung adenocarcinomas<sup>108-109</sup>. Expression of cyclin D1 has been reported in 59% (36/61) of

non-small cell lung cancer tumors analyzed but was not reported to be associated with clinicopathologic parameters<sup>110</sup>.

**FINDING SUMMARY**

CCND1 encodes cyclin D1, a binding partner of the kinases CDK4 and CDK6, that regulates RB activity and cell cycle progression. Amplification of CCND1 has been positively correlated with cyclin D1 overexpression<sup>111</sup> and may lead to excessive proliferation<sup>112-113</sup>.

**GENE**  
**FGF19**
**ALTERATION**  
 amplification

**POTENTIAL TREATMENT STRATEGIES**

There are no targeted therapies that directly address genomic alterations in FGF19. However, amplification of FGF19 predicts sensitivity to inhibitors of FGFR4 in liver cancer cell lines<sup>114</sup>; in one preclinical study, selective inhibition of FGFR4 reduced tumor burden in an FGF19-amplified HCC xenograft model<sup>115</sup>. A Phase 1 study of the FGFR4 inhibitor BLU-554 for previously treated HCC (11/14 sorafenib) reported 1 partial response and 1 stable disease (SD) in patients with FGF19-positive HCC<sup>116</sup>. Preliminary results from the dose escalation part of a Phase 1/2 study evaluating another FGFR4 inhibitor,

FGF4<sup>01</sup>, showed an overall response rate of 8% (4/53), 53% (28/53) SDs, and a median time to progression of 4.1 months; responses were observed in both FGF19-positive and -negative cases<sup>117</sup>. In one clinical study, a trend toward response to sorafenib treatment and FGF19 copy number gain was observed in patients with HCC, and 2 patients harboring FGF19 copy number gain experienced a complete response<sup>118</sup>. Multiple therapies targeting FGF19 or FGFR4 signaling are in preclinical development<sup>119</sup>, and clinical trials evaluating inhibitors of FGFR4 are under way for patients with solid tumors.

**FREQUENCY & PROGNOSIS**

In the TCGA datasets, FGF19 amplification has been reported with highest incidence in esophageal carcinoma (35%), head and neck squamous cell carcinoma (28%), breast carcinoma (16%), lung squamous cell carcinoma (12%), bladder urothelial carcinoma (12%), and cholangiocarcinoma (11%) (cBioPortal, 2017). In HCC, FGF19 is an

important driver gene<sup>115, 120-121</sup>, and FGF19 protein expression correlates with tumor progression and poorer prognosis<sup>122</sup>. Exogenous FGF19 has been shown to promote prostate cancer tumorigenesis in a preclinical study<sup>123</sup>, and the presence of FGF19-positive tissues is an independent factor for worse prognosis following radical prostatectomy<sup>124</sup>.

**FINDING SUMMARY**

FGF19 encodes fibroblast growth factor 19, an FGFR4 ligand involved with bile acid synthesis and hepatocyte proliferation in the liver<sup>115, 125</sup>. FGF19 lies in a region of chromosome 11q13 frequently amplified in a diverse range of malignancies that also contains FGF3, FGF4, and CCND1<sup>126</sup>. Correlation between FGF19 amplification and protein expression has been demonstrated in hepatocellular carcinoma (HCC)<sup>127</sup> but was not observed in several other tumor types<sup>120</sup>.

**GENE**  
**FGF3**
**ALTERATION**  
 amplification

**POTENTIAL TREATMENT STRATEGIES**

There are no targeted therapies that directly address genomic alterations in FGF3. Inhibitors of FGF receptors, however, are

undergoing clinical trials in a number of different cancers.

**FREQUENCY & PROGNOSIS**

FGF3 lies in a region of chromosome 11q13 that also contains FGF19, FGF4, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell cycle progression. This chromosomal region is frequently amplified in a diverse range of malignancies<sup>112</sup>.

**FINDING SUMMARY**

FGF3 encodes fibroblast growth factor 3, a growth factor that plays a central role in

development of the inner ear. Germline mutations in FGF3 give rise to an autosomal recessive syndrome characterized by microdontia, deafness, and complete lack of inner ear structures<sup>128</sup>.

QRF# XXXXXXXX

**GENOMIC FINDINGS**
**GENE**  
**FGF4**
**ALTERATION**  
 amplification

**POTENTIAL TREATMENT STRATEGIES**

FGF4 amplification and overexpression was associated with cell sensitivity to the multikinase inhibitor sorafenib in preclinical studies<sup>129-130</sup> and amplification of FGF4/FGF3 in HCC significantly correlated with patient response to sorafenib ( $p=0.006$ )<sup>129</sup>. Therefore, thyroid carcinoma. Sorafenib is under

investigation in clinical trials in multiple tumor types. FGF4 amplification may confer sensitivity to sorafenib, which is FDA approved to treat HCC, renal cell carcinoma, and differentiated

**FREQUENCY & PROGNOSIS**

This chromosomal region is frequently amplified in a diverse range of malignancies<sup>112</sup> including esophageal carcinoma (35%), head and neck squamous cell carcinoma (HNSCC; 28%), breast invasive carcinoma (16%), lung squamous cell carcinoma (12%), bladder urothelial carcinoma (12%), ovarian serous cystadenocarcinoma (8%), stomach adenocarcinoma (7%), skin melanoma (6%), and hepatocellular carcinoma (HCC; 5%) (cBioPortal, 2017).

**FINDING SUMMARY**

FGF4 encodes fibroblast growth factor 4, which plays a central role in development of the teeth<sup>131</sup> and acts synergistically with other FGFs and SHH (sonic hedgehog) to regulate limb outgrowth in vertebrate development<sup>132</sup>. FGF4 lies in a region of chromosome 11q13 that also contains FGF19, FGF3, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell cycle progression. Amplification of FGF4, along with that of FGF3, FGF19, and CCND1, has been reported in a variety of cancers<sup>112,129,133-136</sup> and may confer sensitivity to the multi-kinase inhibitor sorafenib<sup>129</sup>.

**GENE**  
**NFKBIA**
**ALTERATION**  
 amplification

**POTENTIAL TREATMENT STRATEGIES**

There are no therapies that directly target NFKBIA amplification or expression.

**FREQUENCY & PROGNOSIS**

In the TCGA datasets, amplification of NFKBIA has been reported with the highest incidence in lung adenocarcinoma (11.7%)<sup>107</sup>, esophageal carcinoma (3.8%), uterine carcinosarcoma (3.6%), lung squamous cell carcinoma (3.4%), and ovarian serous cystadenocarcinoma (2.6%) (cBioPortal, 2017). Amplification or increased expression of NFKBIA in EGFR-mutant lung cancer has been reported to predict improved response to EGFR tyrosine kinase inhibitors<sup>137-138</sup>. Certain NFKBIA polymorphisms, which may affect IkBa expression levels, have been studied as

risk factors for some cancer types, although the data are mixed and conflicting<sup>139-141</sup>.

**FINDING SUMMARY**

NFKBIA encodes IkBa, an inhibitor of the NF-kappaB (Nfkb)/REL complex. It has been reported to act as a tumor suppressor in Hodgkin's lymphoma<sup>395-399</sup> and in glioblastoma<sup>392,400-401</sup>. NFKBIA has been reported to be amplified in cancer<sup>227</sup> and may be biologically relevant in this context<sup>228-229</sup>. In contrast, truncating mutations that result in loss of the majority of the IkBa protein are predicted to be inactivating.

**GENE**  
**NKX2-1**
**ALTERATION**  
 amplification

**POTENTIAL TREATMENT STRATEGIES**

There are no approved therapies or trials that target tumors with TTF-1 amplification or overexpression. Lung cancer cell lines that express both TTF-1 and NKX2-8, which is located in the same amplicon as NKX2-1, have demonstrated resistance to cisplatin therapy<sup>152</sup>,

although conflicting data has also been reported<sup>153</sup>.

**FREQUENCY & PROGNOSIS**

Putative amplification of NKX2-1 has been reported with the highest incidence in lung cancer, and has been observed in 14% of adenocarcinomas<sup>107</sup> and 5% of squamous cell carcinomas (SCC)<sup>154</sup> as well as other tumor types including prostate adenocarcinomas (6%)<sup>155</sup>, and poorly differentiated and anaplastic thyroid cancers (4%)<sup>156</sup>. NKX2-1 mutation has been observed in 9% of acinar cell carcinomas of the pancreas<sup>157</sup>, 5% of uterine carcinosarcomas<sup>158</sup>, and is infrequent in other tumor types (cBioPortal, COSMIC, 2018). TTF-1 is expressed in a majority of lung adenocarcinomas and small cell carcinomas, as

well as in a subset of thyroid and CNS tumors<sup>159-161</sup>. Cytoplasmic TTF-1 expression has been reported as an adverse prognostic factor in breast carcinoma<sup>162-163</sup>. However, whether amplification and/or expression status of NKX2-1 have prognostic implications for patients with lung cancer is controversial<sup>152-153,164-167</sup>. TTF-1 has been observed to have tumor-promoting as well as anti-oncogenic roles<sup>168-169</sup>.

**FINDING SUMMARY**

NKX2-1 (NK2 homeobox 1) encodes the thyroid transcription factor TTF-1<sup>170</sup>. Amplification of NKX2-1 results in overexpression of TTF-1 and upregulated transcription of downstream target genes<sup>171</sup>.



QRF# XXXXXXXX

**GENOMIC FINDINGS**
**GENE**  
**TP53**
**ALTERATION**  
**R306\***
**POTENTIAL TREATMENT STRATEGIES**

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor AZD1775<sup>172-175</sup> or p53 gene therapy and immunotherapeutics such as SGT-53<sup>176-180</sup> and ALT-801<sup>181</sup>. In a Phase 1 study, AZD1775 in combination with gemcitabine, cisplatin, or carboplatin elicited partial response in 10% (17/176) and stable disease in 53% (94/176) of patients with solid tumors; the response rate was 21% (4/19) in patients with TP53 mutations versus 12% (4/33) in patients who were TP53-wild-type<sup>182</sup>. Combination of AZD1775 with paclitaxel and carboplatin achieved significantly longer progression-free survival than paclitaxel and carboplatin alone in patients with TP53-mutant ovarian cancer<sup>183</sup>. Furthermore,

AZD1775 in combination with carboplatin achieved a 27% (6/22) response rate and 41% (9/22) stable disease rate in patients with TP53-mutant ovarian cancer refractory or resistant to carboplatin plus paclitaxel<sup>184</sup>. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel in patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including two confirmed and one unconfirmed partial responses and two instances of stable disease with significant tumor shrinkage<sup>180</sup>. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53 mutant, but not TP53 wild-type, breast cancer xenotransplant mouse model<sup>185</sup>. Clinical trials of these agents are under way for some tumor types for patients with a TP53 mutation.

**FREQUENCY & PROGNOSIS**

TP53 is one of the most commonly mutated genes in lung cancer. TP53 mutations have been reported in 43-80% of non-small cell lung cancers (NSCLCs)<sup>107,154,186-191</sup>. Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma<sup>192</sup>. In one study of 55 patients with lung adenocarcinoma, TP53 alterations

correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to PD-1 inhibitors pembrolizumab and nivolumab in this study<sup>24</sup>.

**FINDING SUMMARY**

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers<sup>193</sup>. Any alteration that results in the disruption or partial or complete loss of the region encoding the TP53 DNA-binding domain (DBD, aa 100-292) or the tetramerization domain (aa 325-356), such as observed here, is thought to dysregulate the transactivation of p53-dependent genes and is predicted to promote tumorigenesis<sup>194-196</sup>. Germline mutations in TP53 are associated with the very rare disorder Li-Fraumeni syndrome and the early onset of many cancers<sup>197-202</sup>. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000<sup>203</sup> to 1:20,000<sup>202</sup>, and in the appropriate clinical context, germline testing of TP53 is recommended.



QRF# XXXXXXXX

**THERAPIES WITH CLINICAL BENEFIT**
**IN PATIENT'S TUMOR TYPE**

## Alectinib

*Assay findings associations*
**ALK**

EML4-ALK fusion (Variant 1)

**AREAS OF THERAPEUTIC USE**

Alectinib is a tyrosine kinase inhibitor that targets ALK and RET and is FDA approved to treat patients with ALK positive, metastatic non-small cell lung cancer (NSCLC).

**GENE ASSOCIATION**

Activating ALK alterations may predict sensitivity to alectinib on the basis of extensive clinical evidence in ALK-rearranged NSCLC<sup>51,20456,205-206</sup>.

**SUPPORTING DATA**

Alectinib has been primarily studied for the treatment of ALK-rearranged NSCLC. In the Phase 3 ALEX study comparing alectinib with crizotinib in ALK-rearranged, inhibitor-naïve NSCLC, patients treated with alectinib experienced significantly improved progression-free survival (PFS), 68.4% versus 48.7% (hazard ratio [HR]=0.47); median PFS was not reached in the alectinib arm and was 11.1 months in the crizotinib arm; and median overall survival (OS) was not reached in either arm at 2 years<sup>207</sup>. Similar results have been reported in the J-ALEX trial for inhibitor-naïve Japanese patients with ALK-positive NSCLC<sup>208</sup>. Alectinib combined with atezolizumab led to an objective response rate (ORR) of 81% (17/21) as first-line treatment for PD-L1 unselected, ALK+ NSCLC<sup>65</sup>. In the context of crizotinib resistance, the Phase 3 ALUR trial for patients with ALK+ NSCLC

reported that alectinib significantly improved PFS relative to chemotherapy (7.1 vs. 1.6 months; HR=0.32)<sup>209</sup>. Phase 1/2 and Phase 2 trials of alectinib in ALK-rearranged NSCLC refractory to crizotinib reported ORRs of 45-55%<sup>56,206,210</sup>, with a reported median duration of response of 11.2-17 months<sup>56,210-211</sup>. Alectinib has demonstrated significant activity against central nervous system (CNS) metastases, such as leptomeningeal metastases, for patients with NSCLC<sup>56,204-207,210,212-216</sup>. In the ALUR trial, alectinib significantly improved ORR for CNS metastases relative to chemotherapy (54.2% vs. 0%)<sup>209</sup>. In the ALEX study, alectinib showed superior efficacy in CNS compared with crizotinib, with 12-month progression rate with CNS disease of 41.4% versus 9.4% and median duration of response in patients with CNS disease at baseline for 17.3 months versus 5.5 months<sup>207</sup>. A Phase 2 study of alectinib for crizotinib-refractory, ALK rearranged NSCLC reported 27% of patients achieving a CNS-specific CR, and an overall CNS disease control rate of 83% (95% confidence interval, 74% to 91%)<sup>56</sup>. In a preliminary study of alectinib in four cases of metastatic, RET-rearranged NSCLC, three of whom had previously been treated with cabozantinib, PRs were observed in two patients (one confirmed and one unconfirmed), with an additional patient exhibiting SD for 6 weeks and one case of progressive disease; improvement in CNS disease was observed in one patient after dose increase<sup>217</sup>.

## Brigatinib

*Assay findings associations*
**ALK**

EML4-ALK fusion (Variant 1)

**AREAS OF THERAPEUTIC USE**

Brigatinib is a kinase inhibitor that targets ALK, ROS1, and mutant EGFR and is FDA approved to treat patients with metastatic anaplastic lymphoma kinase (ALK)-positive non-small cell lung cancer (NSCLC) who have progressed on or are intolerant to crizotinib.

**GENE ASSOCIATION**

Activating ALK alterations may predict sensitivity to brigatinib based on strong clinical<sup>218-219</sup> and preclinical<sup>220-221</sup> evidence.

**SUPPORTING DATA**

Brigatinib has been studied primarily for the treatment of ALK-rearranged NSCLC. In the randomized Phase 2 ALTA study, 222 patients with ALK-rearranged NSCLC who progressed on crizotinib were treated with brigatinib and experienced overall response rates (ORRs)

of 48-53% (with 5 CR, 4 PR) and progression-free survival (PFS) rates of 9.2-15.6 months (hazard ratio of 0.55)<sup>219,222</sup>. In addition, brigatinib demonstrated activity against brain metastasis of patients with ALK-rearranged NSCLC, with 23% (18/79; 2 CR, 7 PR) of patients in the 90 mg dose arm achieving a mean intracranial PFS of 15.6 months (hazard ratio of 0.66), although the intracranial PFS was not reached in 18% (13/72, 12 PR) of patients in the 180 mg dose arm of the study<sup>222</sup>. In the expansion stage of a Phase 1/2 study, responses to brigatinib were observed in ALK-rearranged NSCLC cases that were ALK-inhibitor naïve (4/4 patients, 100% ORR or previously treated with crizotinib (31/42 patients, 74% ORR) but not in the single case of EGFR T790M-positive NSCLC with resistance to previous EGFR tyrosine kinase inhibitor<sup>53</sup>. Brigatinib was associated with an ORR of 17% (3/18 patients) in other solid tumors with ALK/ROS1/EGFR alterations<sup>53</sup>.

QRF# XXXXXXXX

**THERAPIES WITH CLINICAL BENEFIT**
**IN PATIENT'S TUMOR TYPE**

## Ceritinib

*Assay findings associations*
**ALK**

EML4-ALK fusion (Variant 1)

**AREAS OF THERAPEUTIC USE**

Ceritinib is an inhibitor of the kinases ALK, ROS1, IR, and IGF-1R. It is FDA approved to treat metastatic nonsmall cell lung cancer (NSCLC) in patients whose tumors are positive for ALK rearrangements, as detected by an FDA-approved test.

**GENE ASSOCIATION**

On the basis of strong clinical data demonstrating benefit to patients with crizotinib-naïve lung cancer<sup>62,223-224</sup> or those previously treated with crizotinib<sup>225-226</sup>, ALK rearrangements may predict sensitivity to ceritinib.

**SUPPORTING DATA**

Multiple Phase 3 studies have reported clinical benefit from ceritinib for patients with advanced ALK-rearranged (ALK+) NSCLC. As a first-line treatment for patients with ALK+ NSCLC in the ASCEND-4 Phase 3 study, ceritinib monotherapy significantly increased the median progression-free survival (PFS) to 16.6 months, compared to a median PFS of 8.1 months in patients with platinum-based chemotherapy<sup>224</sup>. A Phase 3 study of ceritinib for ALK inhibitor-naïve patients with ALK+

NSCLC observed a whole-body (WB) objective response rate (ORR) of 63.7%, a WB disease control rate (DCR) of 89.5%, and progression-free survival (PFS) of 11.1 months<sup>223</sup>. The ASCEND-5 Phase 3 study comparing ceritinib to chemotherapy for patients with ALK+ NSCLC previously treated with crizotinib and chemotherapy also reported a significant benefit for ceritinib in ORR (39% vs. 7%) and median PFS (5.4 vs. 1.6 months); there was no improvement of median OS (18.1 vs. 20.1 months), which may be due to the crossover of patients to the ceritinib arm<sup>226</sup>. The ASCEND-1 Phase 1 study of ceritinib for patients with ALK+ NSCLC reported an ORR of 72%, median PFS of 18.4 months, and 12-month overall survival (OS) of 83%<sup>62</sup>. Earlier Phase 1 and 2 studies reported similar clinical benefit as measured by ORR (39-57%), median PFS (5.7-6.9 months), and median OS of 16.7 months<sup>55,62,227</sup>; for patients with brain metastases, an intracranial ORR of 39% and duration of response of 12.8 months were achieved<sup>225</sup>. Case studies have also reported responses to ceritinib in patients with ALK+ NSCLC and ALK missense mutation after disease progression on crizotinib<sup>228</sup> or alectinib<sup>229-230</sup>.

## Crizotinib

*Assay findings associations*
**ALK**

EML4-ALK fusion (Variant 1)

**AREAS OF THERAPEUTIC USE**

Crizotinib is an inhibitor of the kinases MET, ALK, ROS1, and RON. It is FDA approved to treat patients with metastatic non-small cell lung cancer (NSCLC) whose tumors are positive for ALK rearrangements or ROS1 rearrangements.

**GENE ASSOCIATION**

ALK activation may predict sensitivity to crizotinib. In patients with ALK-rearranged NSCLC, crizotinib improved outcomes in both the first-line<sup>231-232</sup> and second-line<sup>54</sup> settings compared with chemotherapy. Retrospective analysis of 35 patients with NSCLC indicated that compared with other EML4-ALK variants, EML4-ALK variant 1 was an independent predictor of improved median PFS (11.0 vs. 4.2 months, hazard ratio of 0.35) on crizotinib treatment<sup>97</sup>. ALK inhibitors have also demonstrated clinical activity in the context of several other cancer types with activating ALK alterations, including thyroid carcinoma, inflammatory myofibroblastic tumors, and anaplastic large cell lymphoma<sup>58,233-234</sup>.

**SUPPORTING DATA**

The Phase 3 PROFILE 1014 study for patients with ALK positive non-squamous NSCLC reported significantly prolonged progression-free survival [PFS, 10.9 vs. 7.0 months, hazard ratio (HR) 0.45] and higher objective response rate (ORR, 74% vs. 45%) with first-line crizotinib compared with pemetrexed and cisplatin or carboplatin<sup>232</sup>. A similar Phase 3 study for East Asian patients confirmed that crizotinib is superior to chemotherapy in this setting (PFS of 11.1 vs. 6.8 months, HR 0.40; ORR of 87.5% vs. 45.6%)<sup>231</sup>. In the ongoing

Phase 3 PROFILE 1007 study for patients with ALK-positive advanced NSCLC and prior platinum-based therapy (NCT00932893), crizotinib significantly improved median PFS (7.7 months vs. 3.0 months), ORR (65% vs. 20%), and quality of life as compared with chemotherapy<sup>54,235</sup>. The three Phase 3 studies observed numerical, but not statistically significant, improvement of overall survival (OS) with crizotinib (HR of 0.82-0.90), although most patients (70-89%) crossed over from the chemotherapy groups to crizotinib treatment<sup>231,236</sup>. The efficacy of crizotinib in patients with brain metastases has also been examined. Prospective comparison of the intracranial efficacy in patients with stable treated brain metastases included in PROFILE 1014 reported significantly prolonged intracranial disease control rate (DCR) at 24 weeks (56% vs. 25%) and PFS (9.0 vs. 4.0 months, HR 0.40) for patients treated with first-line crizotinib as compared with chemotherapy<sup>237</sup>. Pooled retrospective analysis of patients with ALK-rearranged NSCLC and concurrent brain metastases from the PROFILE 1007 and 1005 studies reported 12-week intracranial DCRs of 56% vs. 62% and intracranial ORR of 18% vs. 33% in patients with previously untreated versus previously treated brain metastases<sup>238</sup>. In a retrospective study of patients with brain metastases from ALK rearranged NSCLC, the majority of whom were treated with radiotherapy and crizotinib, the median OS after diagnosis of brain metastasis was 49.5 months; lack of prior targeted therapy, absence of extracranial metastasis, and a Karnofsky performance score of 90 or higher were significantly associated with improved OS<sup>239</sup>. Upon disease progression, further survival benefit can be derived for patients with ALK-positive NSCLC who continue crizotinib treatment<sup>240</sup>.

QRF# XXXXXXXX

## THERAPIES WITH CLINICAL BENEFIT

## IN OTHER TUMOR TYPE

## Abemaciclib

*Assay findings associations*
**CCND1**  
amplification

### AREAS OF THERAPEUTIC USE

Abemaciclib inhibits the cyclin-dependent kinases 4 and 6 (CDK4/6) and is FDA approved to treat hormone receptor-positive (HR+), HER2-negative (HER2-) advanced or metastatic breast cancer in combination with an aromatase inhibitor as initial endocrine-based therapy for postmenopausal women, in combination with fulvestrant for women who have progressed on endocrine therapy, or as monotherapy for adults who have progressed on endocrine therapy and chemotherapy in the metastatic setting.

### GENE ASSOCIATION

On the basis of clinical data in breast cancer and mantle cell lymphoma<sup>101,105</sup>, CCND1 amplification or activation

may be associated with response to abemaciclib. In a Phase 1 study, 4/10 patients with CCND1-amplified breast cancer responded to single-agent abemaciclib, with all of the responders having HR+ tumors<sup>105</sup>.

### SUPPORTING DATA

Abemaciclib has been investigated primarily in the context of breast cancer<sup>105,241-242</sup>. In a Phase 1 study evaluating abemaciclib as monotherapy, patients with NSCLC experienced a disease control rate of 49% (39% for KRAS wild-type tumors and 55% for KRAS-mutant tumors), with 2 partial responses (PRs)<sup>105</sup>. A Phase 1 study of abemaciclib in combination with ramucirumab in metastatic NSCLC reported 2 unconfirmed PRs<sup>243</sup>.

## Palbociclib

*Assay findings associations*
**CCND1**  
amplification

### AREAS OF THERAPEUTIC USE

Palbociclib inhibits the cyclin-dependent kinases 4 and 6 (CDK4/6) and is FDA approved to treat hormone receptor (HR)-positive/HER2-negative advanced or metastatic breast cancer in combination with an aromatase inhibitor as first-line therapy for postmenopausal women or in combination with fulvestrant following progression on endocrine therapy.

### GENE ASSOCIATION

Clinical studies in liposarcoma and mantle cell lymphoma as well as responses in patients with breast cancer or melanoma indicate that activation of cyclin D-CDK4/6

may predict sensitivity to therapies such as palbociclib<sup>99,106,244</sup>.

### SUPPORTING DATA

Palbociclib has been studied primarily for the treatment of ER+ breast cancer<sup>103,245-246</sup>. A Phase 2 study of palbociclib in patients with recurrent or metastatic nonsmall cell lung cancer (NSCLC) and loss of p16INK4a reported no responses in any of the 16 evaluable patients but stable disease (SD) in 8 (50%) patients<sup>247</sup>. A trial of the CDK4/6 inhibitor abemaciclib in patients with NSCLC reported a disease control rate of 51% (37% for patients with KRAS wild-type tumors and 54% for patients with KRAS-mutant tumors), with one confirmed PR<sup>248</sup>.

## Ribociclib

*Assay findings associations*
**CCND1**  
amplification

### AREAS OF THERAPEUTIC USE

Ribociclib inhibits the cyclin-dependent kinases 4 and 6 (CDK4/6) and is FDA approved in combination with an aromatase inhibitor as first-line therapy to treat women with hormone receptor-positive (HR+), human epidermal growth factor receptor 2-negative (HER2-) advanced or metastatic breast cancer. Ribociclib is also approved in combination with fulvestrant to treat postmenopausal women with HR+, HER2- advanced or metastatic breast cancer, either as first-line therapy or following disease progression on endocrine therapy.

### GENE ASSOCIATION

On the basis of clinical responses for 3 patients with bladder cancer, BRAF/NRAS-wild-type melanoma, or ER positive breast cancer<sup>99,104</sup>, CCND1 amplification may predict sensitivity to CDK4/6 inhibitors such as ribociclib. In a prospective trial, 1 out of 12 patients with CCND1-amplified solid tumors responded to ribociclib<sup>99</sup>.

### SUPPORTING DATA

The Phase 1 Signature study of ribociclib for the treatment of patients with CDK4/6 pathway activated tumors reported clinical benefit for 18.4% (19/103) of cases, 58% (11/19) of whom had p16INK4a mutation or loss; antitumor activity was observed in 3 patients<sup>99</sup>. Phase 1 studies of ribociclib for the treatment of patients with Rb+ advanced solid tumors reported 2.4% partial responses and 23.5-34.4% stable diseases (SD)<sup>104,249</sup>; the 3 responders had alterations in the CDK4/6 pathway<sup>104</sup>. Another Phase 1 study of ribociclib monotherapy reported some efficacy in pediatric patients with neuroblastoma [4 SD, including 2 for >280 days, and 4 progressive disease (PD)] and CNS rhabdoid tumors, including ATRT [1 SD (ongoing after 444 days) and 9 PD], although RB1 status was not determined in any of the patients; of the patients with CDK4-amplified tumors (all neuroblastoma), 1 achieved SD (for >280 days) and 2 exhibited PD<sup>250</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain FDA approved drugs, however the agents listed in this report may have little or no evidence in the patient's tumor type.

QRF# XXXXXXXX

**CLINICAL TRIALS**

**IMPORTANT** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

**GENE**  
**ALK**
**ALTERATION**
**EML4-ALK fusion (Variant 1)**
**RATIONALE**

ALK rearrangements, activating mutations, or amplification may be associated with increased activity in the ALK kinase. Therefore, drugs that inhibit ALK kinase may be relevant. Additionally, patients who have become resistant to crizotinib may harbor sensitivity to newer ALK inhibitors or to HSP90 inhibitors. Examples of clinical trials that may be appropriate for this patient are listed

below. These trials were identified through a search of the trial website [clinicaltrials.gov](https://www.clinicaltrials.gov) using keyword terms such as "alectinib", "AF802", "CH5424802", "ceritinib", "LDK378", "crizotinib", "PF-02341066", "CEP-37440", "dalantercept", "gilteritinib", "ASP2215", "PF-06463922", "RXDX-101", "TSR-011", "X-396", "lung", "solid tumor", and/or "advanced cancer".

**NCT03178552**
**PHASE 2 / 3**

A Phase II/III Multicenter Study Evaluating the Efficacy and Safety of Multiple Targeted Therapies as Treatments for Patients With Advanced or Metastatic Non-Small Cell Lung Cancer (NSCLC) Harboring Actionable Somatic Mutations Detected in Blood (B-FAST: Blood-First Assay Screening Trial)

**TARGETS**  
**ALK, PD-L1, RET**

**LOCATIONS:** Okayama (Japan), Shizuoka (Japan), Saga (Japan), Aichi (Japan), Hiroshima (Japan), Kurralta Park (Australia), Rio de Janeiro (Brazil), California, Krakow (Poland), Moscovskaya Oblast (Russian Federation), CD Mexico (Mexico), Kyoto (Japan), Malaga (Spain), Connecticut, San Luis Potosí (Mexico), Miyagi (Japan), Gdańsk (Poland), Santiago de Compostela (Spain), Warszawa (Poland), Madrid (Spain), Osaka (Japan), Esslingen (Germany), Bunkyo-ku (Japan), Ijuí (Brazil), Ishikawa (Japan), Yamaguchi (Japan), Alicante (Spain), Barcelona (Spain), Shatin (Hong Kong), Hospitalet de Llobregat (Spain), Poitiers (France), Pennsylvania, Tokyo (Japan), Valencia (Spain), Toronto (Canada), Fukuoka (Japan), New York, Wakayama (Japan), Milano (Italy), Beer Sheva (Israel), Olsztyn (Poland), Florida, Illinois, Niigata (Japan), Ehime (Japan), Kanagawa (Japan), Otwock (Poland)

**NCT02767804**
**PHASE 3**

Phase 3 Randomized Study Comparing X-396 (Emsartinib) to Crizotinib in Anaplastic Lymphoma Kinase (ALK) Positive Non-Small Cell Lung Cancer (NSCLC) Patients

**TARGETS**  
**ABL, MET, ALK, ROS1, AXL, TRKC, TRKA**

**LOCATIONS:** Pergamino (Argentina), Virginia, Changchun (China), Barcelona (Spain), Wisconsin, Nanchang (China), São Paulo (Brazil), Changsha (China), Bristol (United Kingdom), Santo André (Brazil), Jerusalem (Israel), Tianjin (China), New York, Warsaw (Poland), Rosario (Argentina), Oregon, Florida, Montpellier (France), Palma de Mallorca (Spain), Edirne (Turkey), Sondrio (Italy), Shenyang (China), Plesice (Czechia), Brussels (Belgium), Ostrava-Vitkovice (Czechia), Gdańsk (Poland), Hong Kong (Hong Kong), Haifa (Israel), Nottingham (United Kingdom), Qingdao (China), Moscow (Russian Federation), Hangzhou (China), Ravenna (Italy), Aviano (Italy), Missouri, Tennessee, Meldola (Italy), Nanjing (China), Idaho, Georgia, Hefei (China), Istanbul (Turkey), Legnago (Italy), Berlin (Germany), Usti nad Labem (Czechia), Beijing (China), Omsk (Russian Federation), Guangzhou (China), Buenos Aires (Argentina), Michigan, Milano (Italy), Lima (Peru), Saint Petersburg (Russian Federation), Pamplona (Spain), Madrid (Spain), Wuhan (China), Izmir (Turkey), Seoul (Korea, Republic of), Caba (Argentina)

**NCT03093116**
**PHASE 1 / 2**

A Phase 1/2, Open-Label, Multi-Center, First-in-Human Study of the Safety, Tolerability, Pharmacokinetics, and Anti-Tumor Activity of TPX-0005 in Patients With Advanced Solid Tumors Harboring ALK, ROS1, or NTRK1-3 Rearrangements (TRIDENT-1)

**TARGETS**  
**ALK, ROS1, TRKC, TRKB, TRKA**

**LOCATIONS:** Massachusetts, Colorado, New York, Seoul (Korea, Republic of), California

**NCT00585195**
**PHASE 1**

Phase 1 Safety, Pharmacokinetic And Pharmacodynamic Study Of Pf-02341066, A C-met/Hgfr Selective Tyrosine Kinase Inhibitor, Administered Orally To Patients With Advanced Cancer

**TARGETS**  
**MET, ALK, ROS1, AXL, TRKC, TRKA**

**LOCATIONS:** New York, Michigan, Colorado, Ohio, Pennsylvania, California, Kashiwa (Japan), Nagoya (Japan), Akashi (Japan), Massachusetts, Melbourne (Australia), North Carolina, Seoul (Korea, Republic of), Vermont, Sapporo (Japan), Osakasayama (Japan)

QRF# XXXXXXXX

**CLINICAL TRIALS**
**NCT02568267**
**PHASE 2**

An Open-Label, Multicenter, Global Phase 2 Basket Study of Entrectinib for the Treatment of Patients With Locally Advanced or Metastatic Solid Tumors That Harbor NTRK1/2/3, ROS1, or ALK Gene Rearrangements

**TARGETS**  
 ALK, ROS1, TRKC, TRKB, TRKA

**LOCATIONS:** Hyogo (Japan), London (United Kingdom), Lyon (France), Leiden (Netherlands), Taipei City (Taiwan), Cambridge (United Kingdom), Florida, Toulouse (France), Oklahoma, Kashiwa-shi (Japan), Washington, Lille (France), Michigan, Illinois, Gdansk (Poland), Barcelona (Spain), Ehime (Japan), Wisconsin, Georgia, Taipei (Taiwan), Köln (Germany), Albury (Australia), Maryland, Göttingen (Germany), Genova (Italy), Warszawa (Poland), Utah, North Carolina, Oregon, New Hampshire, Missouri, Padova (Italy), Madrid (Spain), Bedford Park (Australia), Gliwice (Poland), Tainan (Taiwan), Chang Hua (Taiwan), Hawaii, Amsterdam (Netherlands), Torino (Italy), Massachusetts, Orbassano (Italy), Roma (Italy), Arizona, Shatin (Hong Kong), Taichung (Taiwan), Villejuif cedex (France), Singapore (Singapore), Connecticut, Aichi (Japan), Marseille cedex 5 (France), Shizuoka (Japan), Otwock (Poland), Pisa (Italy), Poznań (Poland), Cheongju-si (Korea, Republic of), Candiolo (Italy), Nevada, Kowloon (Hong Kong), Bordeaux (France), Dresden (Germany), Virginia, Paris (France), Napoli (Italy), District of Columbia, Heidelberg (Australia), New Lambton Heights (Australia), Malaga (Spain), Montpellier cedex 5 (France), Berlin (Germany), Colorado, Paris cedex 15 (France), Miyagi (Japan), Texas, California, Liverpool (Australia), Manchester (United Kingdom), Ohio, Sevilla (Spain), Fukuoka (Japan), Osaka (Japan), Minnesota, Marseille (France), Fuenlabrada (Spain), Milano (Italy), Niigata (Japan), Perugia (Italy), New York, Seoul (Korea, Republic of), Hong Kong (Hong Kong)

**NCT02693535**

Targeted Agent and Profiling Utilization Registry (TAPUR) Study

**TARGETS**  
 ABL, CDK4, PARP, EGFR, DDR2, PDGFRs, VEGFRs, CTLA-4, ROS1, CSF1R, ERBB2, PD-1, ERBB3, MEK, RAF1, KIT, AXL, SMO, TRKC, mTOR, TRKA, MET, ALK, BRAF, RET, SRC, FLT3, CDK6

**LOCATIONS:** North Dakota, Washington, Illinois, California, Pennsylvania, Georgia, Arizona, Utah, North Carolina, Oklahoma, Alabama, South Dakota, Florida, Michigan, Oregon, Virginia, Texas, Nebraska

**NCT01625234**
**PHASE 1 / 2**

Phase 1/2, First-in-Human, Dose-Escalation Study of X-396 (Ersartinib) in Patients With Advanced Solid Tumors and Expansion Phase in Patients With ALK-positive Non-Small Cell Lung Cancer

**TARGETS**  
 ABL, MET, ALK, ROS1, AXL

**LOCATIONS:** California, Oregon, Wisconsin, Tennessee, New York, Missouri, Maryland, South Carolina, Pennsylvania, Massachusetts, Texas, Virginia, Ohio, West Virginia

**NCT02706626**
**PHASE 2**

Phase 2 Trial of Brigatinib After Treatment With Second-Generation ALK Inhibitors in Refractory ALK Rearranged Non-Small Cell Lung Cancer (NSCLC)

**TARGETS**  
 EGFR, ALK, ROS1

**LOCATIONS:** Colorado, Tennessee, Texas, North Carolina

**NCT02321501**
**PHASE 1**

A Phase I/Ib Dose Escalation and Biomarker Study of Ceritinib (LDK378) in Combination With Everolimus in Patients With Locally Advanced or Metastatic Solid Tumors With an Expansion in Non-Small Cell Lung Cancer (NSCLC) Characterized by Abnormalities in Anaplastic Lymphoma Kinase (ALK) Expression

**TARGETS**  
 ALK, ROS1, mTOR

**LOCATIONS:** Texas

QRF# XXXXXXXX

CLINICAL TRIALS

**NCT02227940**

**PHASE 1**

A Phase I Study of Ceritinib (LDK378), a Novel ALK Inhibitor, in Combination With Gemcitabine-Based Chemotherapy in Patients With Advanced Solid Tumors

**TARGETS**  
**ALK, ROS1**

**LOCATIONS:** New York



QRF# XXXXXXXX

**CLINICAL TRIALS**

 GENE  
**CCND1**

 ALTERATION  
amplification

**RATIONALE**

CCND1 amplification may activate CDK4/6 and may predict sensitivity to CDK4/6 inhibitors. Examples of clinical trials that may be appropriate for this patient are listed below. These trials were identified through a search of the trial website

clinicaltrials.gov using keyword terms such as "CDK4", "CDK6", "palbociclib", "PD-0332991", "abemaciclib", "LY2835219", "ribociclib", "LEE011", "NSCLC", "lung", "solid tumor", and/or "advanced cancer".

**NCT03099174**
**PHASE 1**

An Open Label, Phase Ib Dose-escalation Study Evaluating the Safety and Tolerability of BI 836845 and Abemaciclib in Patients With Locally Advanced or Metastatic Solid Tumors and in Combination With Endocrine Therapy in Patients With Locally Advanced or Metastatic Hormone Receptor-positive Breast Cancer, Followed by Expansion Cohorts

**TARGETS**

**CDK4, Aromatase, ER, IGF-2, IGF-1, CDK6**

**LOCATIONS:** Nevada, Madrid (Spain), Connecticut, Pozuelo de Alarcón (Spain), Paris (France), Marseille (France), Barcelona (Spain), California, Minnesota

**NCT02897375**
**PHASE 1**

A Phase I Study of Palbociclib in Combination With Cisplatin or Carboplatin in Advanced Solid Malignancies

**TARGETS**

**CDK4, CDK6**

**LOCATIONS:** Georgia

**NCT01037790**
**PHASE 2**

Phase II Trial of the Cyclin-Dependent Kinase Inhibitor PD 0332991 in Patients With Cancer

**TARGETS**

**CDK4, CDK6**

**LOCATIONS:** Pennsylvania

**NCT03065062**
**PHASE 1**

Phase I Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

**TARGETS**

**CDK4, mTORC1, PI3K-gamma, mTORC2, PI3K-alpha, CDK6**

**LOCATIONS:** Massachusetts



QRF# XXXXXXXX

**APPENDIX**

## Variants of Unknown Significance

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**ASXL1**  
N986S

**CREBBP**  
K2075R

**ERBB2**  
E503K

**FANCC**  
C206F

**KDM5C**  
R1435C

**MEN1**  
amplification

**NOTCH1**  
D1953H

**SMARCA4**  
Q347K

QRF# XXXXXXXX

**APPENDIX**

Genes assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	ATRX	AMER1 (FAM123B)
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTB	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MTOR	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NTSC2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2	PARK2
PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD1)	PDCD1LG2 (PD-L2)	PDGFRA	PDGFRB
PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2	POLD1
POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1	PTEN
PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C	RAD51D
RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET	RICTOR
RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2	SF3B1
SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3	SOX2
SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU	SYK
TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1
TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WHSC1L1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	KIT	EGFR
ETV4	ETV5	ETV6	EWRS1	EZR	FGFR1	FGFR2	FGFR3	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPPRSS2

\*TERC is an ncRNA

\*\*Promoter region of TERT is interrogated

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

QRF# XXXXXXXX

**APPENDIX**

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplastraat 3, 2440 Geel, Belgium.


**ABOUT FOUNDATIONONE CDx**

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:  
[www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

**INTENDED USE**

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI) and tumor mutational burden (TMB) using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

**TEST PRINCIPLE**

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes. Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X).

Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g. gene fusions). Additionally, genomic signatures including microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

**THE REPORT**

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

**Diagnostic Significance**

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

**Qualified Alteration Calls (Equivocal and Subclonal)**

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

**Ranking of Alterations and Therapies  
Biomarker Findings**

Appear at the top of the report, but are not ranked higher than Genomic Findings.

**Genomic Findings**

Therapies with Clinical Benefit In Patient's Tumor Type → Therapies with Clinical Benefit in Other Tumor Type → Clinical Trial Options → No Known Options (If multiple findings exist within any of these categories, the results are listed alphabetically by gene name.)

**Therapies**

Sensitizing therapies → Resistant therapies. (If multiple therapies exist within any of these categories, they are listed alphabetically by therapy name.)

**Clinical Trials**

Pediatric trial qualification → Geographical proximity → Later trial phase.

**Limitations**

1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established.
2. TMB by F1CDx is defined based on counting the total number of all synonymous and nonsynonymous variants present at 5% allele frequency or greater (after filtering) and reported as mutations per megabase (mut/Mb) unit rounded to the nearest integer. The clinical validity of TMB defined by this panel has not been established.

QRF# XXXXXXXX

**APPENDIX**

About FoundationOne®CDx

**LEVEL OF EVIDENCE NOT PROVIDED**

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

**NO GUARANTEE OF CLINICAL BENEFIT**

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

**NO GUARANTEE OF REIMBURSEMENT**

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

**TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN**

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

**SELECT ABBREVIATIONS**

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

The median exon coverage for this sample is 733X

**QRF#** XXXXXXXX

<b>APPENDIX</b>	<b>References</b>
-----------------	-------------------

1. Gatalica Z, Snyder C, Maney T, et al. (2014) Programmed cell death 1 (PD-1) and its ligand (PD-L1) in common cancers and their correlation with molecular cancer type. *Cancer Epidemiol Biomarkers Prev* 23(12):2965-70
2. Kroemer G, Galluzzi L, Zitvogel L, et al. (2015) Colorectal cancer: the first neoplasia found to be under immunosurveillance and the last one to respond to immunotherapy? *Oncoimmunology* 4(7):e1058597
3. Lal N, Beggs AD, Willcox BE, et al. (2015) An immunogenomic stratification of colorectal cancer: Implications for development of targeted immunotherapy. *Oncoimmunology* 4(3):e976052
4. Overman et al., 2016; ASCO Abstract 3501
5. Le DT, Uram JN, Wang H, et al. (2015) PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. *N Engl J Med* ePub May 2015
6. ASCO-SITC 2016; Abstract P60
7. Warth A, Körner S, Penzel R, et al. (2016) Microsatellite instability in pulmonary adenocarcinomas: a comprehensive study of 480 cases. *Virchows Arch* 468(3):313-9
8. Ninomiya H, Nomura K, Satoh Y, et al. (2006) Genetic instability in lung cancer: concurrent analysis of chromosomal, mini- and microsatellite instability and loss of heterozygosity. *Br J Cancer* 94(10):1485-91
9. Woenckhaus M, Stoehr R, Dietmaier W, et al. (2003) Microsatellite instability at chromosome 8p in nonsmall cell lung cancer is associated with lymph node metastasis and squamous differentiation. *Int J Oncol* 23(5):1357-63
10. Chang JW, Chen YC, Chen CY, et al. (2000) Correlation of genetic instability with mismatch repair protein expression and p53 mutations in nonsmall cell lung cancer. *Clin Cancer Res* 6(5):1639-46
11. Fong KM, Zimmerman PV, Smith PJ (1995) Microsatellite instability and other molecular abnormalities in non-small cell lung cancer. *Cancer Res* 55(1):28-30
12. Hansen LT, Thykjaer T, Ørntoft TF, et al. (2003) The role of mismatch repair in small-cell lung cancer cells. *Eur J Cancer* 39(10):1456-67
13. Kocarnik JM, Shiovitz S, Phipps AI (2015) Molecular phenotypes of colorectal cancer and potential clinical applications. *Gastroenterol Rep (Oxf)* 3(4):269-76
14. You JF, Buhard O, Ligtenberg MJ, et al. (2010) Tumours with loss of MSH6 expression are MSI-H when screened with a pentaplex of five mononucleotide repeats. *Br J Cancer* 103(12):1840-5
15. Bairwa NK, Saha A, Gochhait S, et al. (2014) Microsatellite instability: an indirect assay to detect defects in the cellular mismatch repair machinery. *Methods Mol Biol* 1105:497-509
16. Boland CR, Thibodeau SN, Hamilton SR, et al. (1998) A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res* 58(22):5248-57
17. Pawlik TM, Raut CP, Rodriguez-Bigas MA (2004) Colorectal carcinogenesis: MSI-H versus MSI-L. *Dis Markers* 20(4-5): 199-206
18. Boland CR, Goel A (2010) Microsatellite instability in colorectal cancer. *Gastroenterology* 138(6):2073-2087.e3
19. Snyder A, Makarov V, Merghoub T, et al. (2014) Genetic basis for clinical response to CTLA-4 blockade in melanoma. *N Engl J Med* 371(23):2189-99
20. Rosenberg JE, Hoffman-Censits J, Powles T, et al. (2016) Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinum-based chemotherapy: a single-arm, multicentre, phase 2 trial. *Lancet* 387(10031):1909-20
21. Johnson DB, Frampton GM, Rieth MJ, et al. (2016) Targeted next generation sequencing identifies markers of response to PD-1 blockade. *Cancer Immunol Res* ePub Sep 2016
22. Balar AV, Galsky MD, Rosenberg JE, et al. (2017) Atezolizumab as first-line treatment in cisplatin-ineligible patients with locally advanced and metastatic urothelial carcinoma: a single-arm, multicentre, phase 2 trial. *Lancet* 389(10064):67-76
23. Miao D, Margolis CA, Vokes NJ, et al. (2018) Genomic correlates of response to immune checkpoint blockade in microsatellite-stable solid tumors. *Nat Genet* 50(9):1271-1281
24. Dong ZY, Zhong WZ, Zhang XC, et al. (2016) Potential Predictive Value of TP53 and KRAS Mutation Status for Response to PD-1 Blockade Immunotherapy in Lung Adenocarcinoma. *Clin Cancer Res*
25. Rizvi NA, Hellmann MD, Snyder A, et al. (2015) Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* 348(6230):124-8
26. Mehnert JM, Panda A, Zhong H, et al. (2016) Immune activation and response to pembrolizumab in POLEmutant endometrial cancer. *J Clin Invest* 126(6):2334-40
27. Santin AD, Bellone S, Buza N, et al. (2016) Regression of chemotherapy-resistant Polymerase epsilon (POLE) ultra-mutated and MSH6 hyper-mutated endometrial tumors with nivolumab. *Clin Cancer Res* ePub Aug 2016
28. Johanns TM, Miller CA, Dorward IG, et al. (2016) Immunogenomics of Hypermutated Glioblastoma: a Patient with Germline POLE Deficiency Treated with Checkpoint Blockade Immunotherapy. *Cancer Discov* ePub Sep 2016
29. Bouffet E, Larouche V, Campbell BB, et al. (2016) Immune Checkpoint Inhibition for Hypermutant Glioblastoma Multiforme Resulting From Germline Biallelic Mismatch Repair Deficiency. *J Clin Oncol* ePub Mar 2016
30. Fabrizio DA, George TJ, Dunne RF, et al. (2018) Beyond microsatellite testing: assessment of tumor mutational burden identifies subsets of colorectal cancer who may respond to immune checkpoint inhibition. *J Gastrointest Oncol* 9(4): 610-617
31. Van Allen EM, Miao D, Schilling B, et al. (2015) Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. *Science* 350(6257):207-11
32. Legrand et al., 2018; ASCO Abstract 12000
33. Spigel et al., 2016; ASCO Abstract 9017
34. Schwartz et al., 2016; ASCO Abstract 8533
35. Xiao D, Pan H, Li F, et al. (2016) Analysis of ultra-deep targeted sequencing reveals mutation burden is associated with gender and clinical outcome in lung adenocarcinoma. *Oncotarget* 7(16):22857-64
36. Shim HS, Kenudson M, Zheng Z, et al. (2015) Unique Genetic and Survival Characteristics of Invasive Mucinous Adenocarcinoma of the Lung. *J Thorac Oncol* 10(8):1156-62
37. Govindan R, Ding L, Griffith M, et al. (2012) Genomic landscape of non-small cell lung cancer in smokers and never-smokers. *Cell* 150(6):1121-34
38. Ding L, Getz G, Wheeler DA, et al. (2008) Somatic mutations affect key pathways in lung adenocarcinoma. *Nature* 455(7216):1069-75
39. Imielinski M, Berger AH, Hammerman PS, et al. (2012) Mapping the hallmarks of lung adenocarcinoma with massively parallel sequencing. *Cell* 150(6):1107-20
40. Kim Y, Hammerman PS, Kim J, et al. (2014) Integrative and comparative genomic analysis of lung squamous cell carcinomas in East Asian patients. *J Clin Oncol* 32(2):121-8
41. Pfeifer GP, You YH, Besaratinia A (2005) Mutations induced by ultraviolet light. *Mutat Res* 571(1-2):19-31
42. Hill VK, Gartner JJ, Samuels Y, et al. (2013) The genetics of melanoma: recent advances. *Annu Rev Genomics Hum Genet* 14:257-79
43. Pfeifer GP, Denissenko MF, Olivier M, et al. (2002) Tobacco smoke carcinogens, DNA damage and p53 mutations in smoking-associated cancers. *Oncogene* 21(48):7435-51
44. Cancer Genome Atlas Research Network, Kandoth C, Schultz N, et al. (2013) Integrated genomic characterization of endometrial carcinoma. *Nature* 497(7447):67-73
45. Briggs S, Tomlinson I (2013) Germline and somatic polymerase ε and δ mutations define a new class of hypermutated colorectal and endometrial cancers. *J Pathol* 230(2):148-53
46. Heitzer E, Tomlinson I (2014) Replicative DNA polymerase mutations in cancer. *Curr Opin Genet Dev* 24:107-13
47. Cancer Genome Atlas Network (2012) Comprehensive molecular characterization of human colon and rectal cancer. *Nature* 487(7407):330-7
48. Roberts SA, Gordenin DA (2014) Hypermutation in human cancer genomes: footprints and mechanisms. *Nat Rev Cancer* 14(12):786-800
49. Camidge et al., 2011; ASCO Abstract 2501
50. Bang et al., 2010; ASCO Abstract 3
51. Gandhi et al., 2015; ASCO Abstract 8019
52. Kwak EL, Bang YJ, Camidge DR, et al. (2010) Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 363(18):1693-703
53. Gettinger SN, Bazhenova LA, Langer CJ, et al. (2016) Activity and safety of brigatinib in ALK-rearranged non-small-cell lung cancer and other malignancies: a single-arm, open-label, phase 1/2 trial. *Lancet Oncol* 17(12):1683-1696
54. Shaw AT, Kim DW, Nakagawa K, et al. (2013) Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med* 368(25):2385-94
55. Shaw AT, Kim DW, Mehra R, et al. (2014) Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med* 370(13): 1189-97
56. Ou SI, Ahn JS, De Petris L, et al. (2015) Alectinib in Crizotinib-Refractory ALK-Rearranged Non-Small-Cell Lung Cancer: A Phase II Global Study. *J Clin Oncol* ePub Nov 2015
57. Solomon BJ, Kim DW, Wu YL, et al. (2018) Final Overall Survival Analysis From a Study Comparing First-Line Crizotinib Versus Chemotherapy in ALK-Mutation-Positive Non-Small-Cell Lung Cancer. *J Clin Oncol* 36(22):2251-2258
58. Mossé YP, Lim MS, Voss SD, et al. (2013) Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: a Children's Oncology Group phase 1 consortium study. *Lancet Oncol* 14(6):472-80
59. Mosse et al., 2012; ASCO 9500
60. Chand D, Yamazaki Y, Ruuth K, et al. (2013) Cell culture and Drosophila model systems define three classes of anaplastic lymphoma kinase mutations in neuroblastoma. *Dis Model Mech* 6(2):373-82
61. Schönherr C, Ruuth K, Yamazaki Y, et al. (2011) Activating ALK mutations found in neuroblastoma are inhibited by Crizotinib and NVP-TAE684. *Biochem J* 440(3):405-13
62. Kim DW, Mehra R, Tan DS, et al. (2016) Activity and safety of ceritinib in patients with ALK-rearranged non-small-cell lung cancer (ASCEND-1): updated results from the multicentre, open-label, phase 1 trial. *Lancet Oncol* ePub Mar 2016
63. Camidge DR, Kim DW, Tiseo M, et al. (2018) Exploratory Analysis of Brigatinib Activity in Patients With Anaplastic Lymphoma Kinase-Positive Non- Small-Cell Lung Cancer and Brain Metastases in Two Clinical Trials. *J Clin Oncol* :JCO2017775841
64. Huber et al., 2018; ASCO Abstract 9061
65. Kim et al., 2018; ASCO Abstract 9009
66. Besse et al., 2018; ASCO Abstract 9032
67. Shaw et al., 2018; AACR Abstract CT044
68. Yoda S, Lin JJ, Lawrence MS, et al. (2018) Sequential ALK Inhibitors Can Select for Lorlatinib-Resistant Compound ALK Mutations in ALK-Positive Lung Cancer. *Cancer Discov* 8(6): 714-729
69. Dagogo-Jack I, Brannon AR, Ferris LA, et al. (2018) Tracking the Evolution of Resistance to ALK Tyrosine Kinase Inhibitors through Longitudinal Analysis of Circulating Tumor DNA. *JCO Precis Oncol* 2018
70. Hochmair MJ, Schwab S, Prosch H (2017) Complete remission of intrathelial metastases with lorlatinib therapy in a heavily pretreated ALK-positive lung cancer patient. *Anticancer Drugs* 28(8):928-930



**QRF# XXXXXXXX**

<b>APPENDIX</b>	<b>References</b>
-----------------	-------------------

71. Yuan C, Ma MJ, Parker JV, et al. (2017) Metastatic Anaplastic Lymphoma Kinase-1 (ALK-1)-Rearranged Inflammatory Myofibroblastic Sarcoma to the Brain with Leptomeningeal Involvement: Favorable Response to Serial ALK Inhibitors: A Case Report. *Am J Case Rep* 18:799-804
72. Shaw et al., 2018; ASCO Abstract 9008
73. Horn L, Infante JR, Reckamp KL, et al. (2018) Ensartinib (X-396) in ALK-Positive Non-Small Cell Lung Cancer: Results from a First-in-Human Phase I/II, Multicenter Study. *Clin Cancer Res* 24(12): 2771-2779
74. Drilon A, Siena S, Ou SI, et al. (2017) Safety and Antitumor Activity of the Multi-Targeted Pan-TRK, ROS1, and ALK Inhibitor Entrectinib (RXDX-101): Combined Results from Two Phase 1 Trials (ALKA-372-001 and STARTRK-1). *Cancer Discov* ePub Feb 2017
75. Socinski MA, Goldman J, El-Hariry I, et al. (2013) A multicenter phase II study of ganetespib monotherapy in patients with genotypically defined advanced non-small cell lung cancer. *Clin Cancer Res* 19(11):3068-77
76. Koivunen JP, Mermel C, Zejnullahu K, et al. (2008) EML4-ALK fusion gene and efficacy of an ALK kinase inhibitor in lung cancer. *Clin Cancer Res* 14(13):4275-83
77. Inamura K, Takeuchi K, Togashi Y, et al. (2009) EML4-ALK lung cancers are characterized by rare other mutations, a TTF-1 cell lineage, an acinar histology, and young onset. *Mod Pathol* 22(4): 508-15
78. Shaw AT, Yeap BY, Mino-Kenudson M, et al. (2009) Clinical features and outcome of patients with non-small-cell lung cancer who harbor EML4-ALK. *J Clin Oncol* 27(26):4247-53
79. Takahashi T, Sonobe M, Kobayashi M, et al. (2010) Clinicopathologic features of non-small-cell lung cancer with EML4-ALK fusion gene. *Ann Surg Oncol* 17(3):889-97
80. Li Y, Li Y, Yang T, et al. (2013) Clinical significance of EML4-ALK fusion gene and association with EGFR and KRAS gene mutations in 208 Chinese patients with non-small cell lung cancer. *PLoS ONE* 8(1):e52093
81. Li H, Pan Y, Li Y, et al. (2013) Frequency of well identified oncogenic driver mutations in lung adenocarcinoma of smokers varies with histological subtypes and graduated smoking dose. *Lung Cancer* 79(1):8-13
82. Zhou JX, Yang H, Deng Q, et al. (2013) Oncogenic driver mutations in patients with non-small-cell lung cancer at various clinical stages. *Ann Oncol* 24(5):1319-25
83. Takeuchi K, Choi YL, Togashi Y, et al. (2009) KIF5BALK, a novel fusion oncoprotein identified by an immunohistochemistry-based diagnostic system for ALK-positive lung cancer. *Clin Cancer Res* 15(9):3143-9
84. To KF, Tong JH, Yeung KS, et al. (2013) Detection of ALK rearrangement by immunohistochemistry in lung adenocarcinoma and the identification of a novel EML4-ALK variant. *J Thorac Oncol* 8(7):883-91
85. Grande E, Bolós MV, Arriola E (2011) Targeting oncogenic ALK: a promising strategy for cancer treatment. *Mol Cancer Ther* 10(4): 569-79
86. Peters S, Taron M, Bubendorf L, et al. (2013) Treatment and detection of ALK-rearranged NSCLC. *Lung Cancer* 81(2):145-54
87. Li T, Maus MK, Desai SJ, et al. (2014) Large-scale screening and molecular characterization of EML4-ALK fusion variants in archival non-small-cell lung cancer tumor specimens using quantitative reverse transcription polymerase chain reaction assays. *J Thorac Oncol* 9(1):18-25
88. Heuckmann JM, Balke-Want H, Malchers F, et al. (2012) Differential protein stability and ALK inhibitor sensitivity of EML4-ALK fusion variants. *Clin Cancer*
89. Lei YY, Yang JJ, Zhang XC, et al. (2015) Anaplastic Lymphoma Kinase Variants and the Percentage of ALK-Positive Tumor Cells and the Efficacy of Crizotinib in Advanced NSCLC. *Clin Lung Cancer* ePub Sep 2015
90. Choi YL, Takeuchi K, Soda M, et al. (2008) Identification of novel isoforms of the EML4-ALK transforming gene in non-small cell lung cancer. *Cancer Res* 68(13):4971-6
91. Sasaki T, Rodig SJ, Chirieac LR, et al. (2010) The biology and treatment of EML4-ALK non-small cell lung cancer. *Eur J Cancer* 46(10):1773-80
92. Takeuchi K, Choi YL, Soda M, et al. (2008) Multiplex reverse transcription-PCR screening for EML4-ALK fusion transcripts. *Clin Cancer Res* 14(20):6618-24
93. Zhang X, Zhang S, Yang X, et al. (2010) Fusion of EML4 and ALK is associated with development of lung adenocarcinomas lacking EGFR and KRAS mutations and is correlated with ALK expression. *Mol Cancer* 9:188
94. Maus MK, Stephens C, Zeger G, et al. (2012) Identification of Novel Variant of EML4-ALK Fusion Gene in NSCLC: Potential Benefits of the RT-PCR Method. *Int J Biomed Sci* 8(1):1-6
95. Sanders HR, Li HR, Bruey JM, et al. (2011) Exon scanning by reverse transcriptase-polymerase chain reaction for detection of known and novel EML4-ALK fusion variants in non-small cell lung cancer. *Cancer Genet* 204(1):45-52
96. Soda M, Choi YL, Enomoto M, et al. (2007) Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 448(7153):561-6
97. Yoshida T, Oya Y, Tanaka K, et al. (2016) Differential Crizotinib Response Duration Among ALK Fusion Variants in ALK-Positive Non-Small-Cell Lung Cancer. *J Clin Oncol* ePub Jun 2016
98. Juric et al., 2016; ASCO Abstract 568
99. Peguero et al., 2016; ASCO Abstract 2528
100. Tolane et al., 2016; SABCS P4-22-12
101. Morschhauser et al., 2014; ASH Abstract 3067
102. Flaherty KT, Lorusso PM, Demichele A, et al. (2012) Phase I, dose-escalation trial of the oral cyclin-dependent kinase 4/6 inhibitor PD 0332991, administered using a 21-day schedule in patients with advanced cancer. *Clin Cancer Res* 18(2):568-76
103. Finn RS, Crown JP, Lang I, et al. (2014) The cyclin-dependent kinase 4/6 inhibitor palbociclib in combination with letrozole versus letrozole alone as first-line treatment of oestrogen receptor-positive, HER2-negative, advanced breast cancer (PALOMA-1/TRIO-18): a randomized phase 2 study. *Lancet Oncol* ePub Dec 2014
104. Infante JR, Cassier PA, Gerecitano JF, et al. (2016) A Phase I Study of the Cyclin-Dependent Kinase 4/6 Inhibitor Ribociclib (LEE011) in Patients with Advanced Solid Tumors and Lymphomas. *Clin Cancer Res* 22(23):5696-5705
105. Patnaik A, Rosen LS, Tolane SM, et al. (2016) Efficacy and Safety of Abemaciclib, an Inhibitor of CDK4 and CDK6, for Patients with Breast Cancer, Non-Small Cell Lung Cancer, and Other Solid Tumors. *Cancer Discov* 6(7):740-53
106. Leonard JP, LaCasce AS, Smith MR, et al. (2012) Selective CDK4/6 inhibition with tumor responses by PD0332991 in patients with mantle cell lymphoma. *Blood* 119(20):4597-607
107. Cancer Genome Atlas Research Network (2014) Comprehensive molecular profiling of lung adenocarcinoma. *Nature* 511(7511):543-50
108. Reissmann PT, Koga H, Figlin RA, et al. (1999) Amplification and overexpression of the cyclin D1 and epidermal growth factor receptor genes in non-small-cell lung cancer. *Lung Cancer Study Group. J Cancer Res Clin Oncol* 125(2):61-70
109. Marchetti A, Dogliani C, Barbareschi M, et al. (1998) Cyclin D1 and retinoblastoma susceptibility gene alterations in non-small cell lung cancer. *Int J Cancer* 75(2):187-92
110. Sun W, Song L, Ai T, et al. (2013) Prognostic value of MET, cyclin D1 and MET gene copy number in non-small cell lung cancer. *J Biomed Res* 27(3):220-30
111. Elsheikh S, Green AR, Aleskandarany MA, et al. (2008) CCND1 amplification and cyclin D1 expression in breast cancer and their relation with proteomic subgroups and patient outcome. *Breast Cancer Res Treat* 109(2):325-35
112. Fu M, Wang C, Li Z, et al. (2004) Minireview: Cyclin D1: normal and abnormal functions. *Endocrinology* 145(12):5439-47
113. Takahashi-Yanaga F, Sasaguri T (2008) GSK-3beta regulates cyclin D1 expression: a new target for chemotherapy. *Cell Signal* 20(4):581-9
114. Guagnano V, Kauffmann A, Wöhrle S, et al. (2012) FGFR genetic alterations predict for sensitivity to NVP-BGJ398, a selective pan-FGFR inhibitor. *Cancer Discov* ePub Sep 2012
115. Hagel M, Miduturu C, Sheets M, et al. (2015) First Selective Small Molecule Inhibitor of FGFR4 for the Treatment of Hepatocellular Carcinomas with an Activated FGFR4 Signaling Pathway. *Cancer Discov* 5(4):424-37
116. Kim et al., 2016; EORTC-NCI-AACR Symposium Abstract 105A
117. Chan et al., 2017; AACR Abstract CT106/24
118. Kaibori M, Sakai K, Ishizaki M, et al. (2016) Increased FGF19 copy number is frequently detected in hepatocellular carcinoma with a complete response after sorafenib treatment. *Oncotarget* ePub Jun 2016
119. Packer LM, Pollock PM (2015) Paralog-Specific Kinase Inhibition of FGFR4: Adding to the Arsenal of Anti-FGFR Agents. *Cancer Discov* 5(4):355-7
120. Sawey ET, Chanion M, Cai C, et al. (2011) Identification of a therapeutic strategy targeting amplified FGF19 in liver cancer by Oncogenic screening. *Cancer Cell* 19(3):347-58
121. Desnoyers LR, Pai R, Ferrando RE, et al. (2008) Targeting FGF19 inhibits tumor growth in colon cancer xenograft and FGF19 transgenic hepatocellular carcinoma models. *Oncogene* 27(1): 85-97
122. Miura S, Mitsunashi N, Shimizu H, et al. (2012) Fibroblast growth factor 19 expression correlates with tumor progression and poorer prognosis of hepatocellular carcinoma. *BMC Cancer* 12:56
123. Feng S, Dakhova O, Creighton CJ, et al. (2013) Endocrine fibroblast growth factor FGF19 promotes prostate cancer progression. *Cancer Res* 73(8):2551-62
124. Nagamatsu H, Teishima J, Goto K, et al. (2015) FGF19 promotes progression of prostate cancer. *Prostate* ePub Apr 2015
125. Xie MH, Holcomb I, Deuel B, et al. (1999) FGF-19, a novel fibroblast growth factor with unique specificity for FGFR4. *Cytokine* 11(10):729-35
126. Katoh M (2002) WNT and FGF gene clusters (review). *Int J Oncol* 21(6):1269-73
127. Kan Z, Zheng H, Liu X, et al. (2013) Whole-genome sequencing identifies recurrent mutations in hepatocellular carcinoma. *Genome Res* 23(9):1422-33
128. Tekin M, Hışmı BO, Fitoz S, et al. (2007) Homozygous mutations in fibroblast growth factor 3 are associated with a new form of syndromic deafness characterized by inner ear agenesis, microtia, and microdontia. *Am J Hum Genet* 80(2): 338-44
129. Arai T, Ueshima K, Matsumoto K, et al. (2013) FGF3/FGF4 amplification and multiple lung metastases in responders to sorafenib in hepatocellular carcinoma. *Hepatology* 57(4): 1407-15
130. Yamada T, Abei M, Danjoh I, et al. (2015) Identification of a unique hepatocellular carcinoma line, Li-7, with CD13(+) cancer stem cells hierarchy and population change upon its differentiation during culture and effects of sorafenib. *BMC Cancer* 15:260
131. Kratochwil K, Galceran J, Tontsch S, et al. (2002) FGF4, a direct target of LEF1 and Wnt signaling, can rescue the arrest of tooth organogenesis in *Lef1(-/-)* mice. *Genes Dev* 16(24):3173-85
132. Scherz PJ, Harfe BD, McMahon AP, et al. (2004) The limb bud *Shh-Fgf* feedback loop is terminated by expansion of former ZPA cells. *Science* 305(5682):396-9
133. Zaharieva BM, Simon R, Diener PA, et al. (2003) Highthroughput tissue microarray analysis of 11q13 gene amplification (CCND1, FGF3, FGF4, EMS1) in urinary bladder cancer. *J Pathol* 201(4):603-8
134. Arai H, Ueno T, Tangoku A, et al. (2003) Detection of amplified oncogenes by genome DNA microarrays in human primary esophageal squamous cell carcinoma: comparison with conventional comparative genomic hybridization analysis. *Cancer Genet Cytogenet* 146(1):16-21

QRF# XXXXXXXX

**APPENDIX**
**References**

135. Ribeiro IP, Marques F, Caramelo F, et al. (2014) Genetic imbalances detected by multiplex ligation-dependent probe amplification in a cohort of patients with oral squamous cell carcinoma—the first step towards clinical personalized medicine. *Tumour Biol* 35(5):4687-95
136. Schulze K, Imbeaud S, Letouzé E, et al. (2015) Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. *Nat Genet* 47(5): 505-11
137. Bivona TG, Hieronymus H, Parker J, et al. (2011) FAS and NF-κB signalling modulate dependence of lung cancers on mutant EGFR. *Nature* 471(7339):523-6
138. Giannikopoulos et al., 2014; ASCO Abstract 8083
139. Zhao Z, Zhong X, Wu T, et al. (2014) Identification of a NFKBIA polymorphism associated with lower NFKBIA protein levels and poor survival outcomes in patients with glioblastoma multiforme. *Int J Mol Med* 34(5):1233-40
140. Geng P, Ou J, Li J, et al. (2015) Genetic Association Between NFKBIA -881A>G Polymorphism and Cancer Susceptibility. *Medicine (Baltimore)* 94(31):e1024
141. Zhang M, Huang J, Tan X, et al. (2015) Common Polymorphisms in the NFKBIA Gene and Cancer Susceptibility: A Meta-Analysis. *Med Sci Monit* 21:3186-96
142. Weniger MA, Küppers R (2016) NF-κB deregulation in Hodgkin lymphoma. *Semin Cancer Biol* ePub May 2016
143. Liu X, Yu H, Yang W, et al. (2010) Mutations of NFKBIA in biopsy specimens from Hodgkin lymphoma. *Cancer Genet Cytogenet* 197(2):152-7
144. Lake A, Shield LA, Cordonato P, et al. (2009) Mutations of NFKBIA, encoding IκappaB alpha, are a recurrent finding in classical Hodgkin lymphoma but are not a unifying feature of non-EBV-associated cases. *Int J Cancer* 125(6):1334-42
145. Birnstiel ML, Jacob J, Sirtlin JL (1965) Analysis of nucleolar RNA synthesis in dipteran salivary glands. *Arch Biol (Liege)* 76(2): 565-89
146. Cabannes E, Khan G, Aillet F, et al. (1999) Mutations in the IκBa gene in Hodgkin's disease suggest a tumour suppressor role for IκappaBalpha. *Oncogene* 18(20):3063-70
147. Bredel M, Scholtens DM, Yadav AK, et al. (2011) NFKBIA deletion in glioblastomas. *N Engl J Med* 364(7):627-37
148. Patané M, Porra P, Bottega E, et al. (2013) Frequency of NFKBIA deletions is low in glioblastomas and skewed in glioblastoma neurospheres. *Mol Cancer* 12:160
149. Gao J, Aksoy BA, Dogrusoz U, et al. (2013) Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal* 6(269):pl1
150. Zack TI, Schumacher SE, Carter SL, et al. (2013) Pancancer patterns of somatic copy number alteration. *Nat Genet* 45(10): 1134-1140
151. Beroukhi R, Mermel CH, Porter D, et al. (2010) The landscape of somatic copy-number alteration across human cancers. *Nature* 463(7283):899-905
152. Hsu DS, Acharya CR, Balakumaran BS, et al. (2009) Characterizing the developmental pathways TTF-1, NKX2-8, and PAX9 in lung cancer. *Proc Natl Acad Sci USA* 106(13):5312-7
153. Yang L, Lin M, Ruan WJ, et al. (2012) Nkx2-1: a novel tumor biomarker of lung cancer. *J Zhejiang Univ Sci B* 13(11):855-66
154. Cancer Genome Atlas Research Network (2012) Comprehensive genomic characterization of squamous cell lung cancers. *Nature* 489(7417):519-25
155. Kumar A, Coleman I, Morrissey C, et al. (2016) Substantial interindividual and limited intraindividual genomic diversity among tumors from men with metastatic prostate cancer. *Nat Med* 22(4):369-78
156. Landa I, Ibrahimipasic T, Boucai L, et al. (2016) Genomic and transcriptomic hallmarks of poorly differentiated and anaplastic thyroid cancers. *J Clin Invest* ePub Feb 2016
157. Jiao Y, Yonescu R, Offerhaus GJ, et al. (2014) Whole-exome sequencing of pancreatic neoplasms with acinar differentiation. *J Pathol* 232(4):428-35
158. Jones S, Stransky N, McCord CL, et al. (2014) Genomic analyses of gynaecologic carcinosarcomas reveal frequent mutations in chromatin remodelling genes. *Nat Commun* 5:5006
159. Nakamura N, Miyagi E, Murata S, et al. (2002) Expression of thyroid transcription factor-1 in normal and neoplastic lung tissues. *Mod Pathol* 15(10):1058-67
160. Moldvay J, Jäckel M, Bogos K, et al. (2004) The role of TTF-1 in differentiating primary and metastatic lung adenocarcinomas. *Pathol Oncol Res* 10(2):85-8
161. Gilbert-Sirieix M, Makoukji J, Kimura S, et al. (2011) Wnt/β-catenin signaling pathway is a direct enhancer of thyroid transcription factor-1 in human papillary thyroid carcinoma cells. *PLoS ONE* 6(7):e22280
162. Robens J, Goldstein L, Gown AM, et al. (2010) Thyroid transcription factor-1 expression in breast carcinomas. *Am J Surg Pathol* 34(12):1881-5
163. Ni YB, Tsang JY, Shao MM, et al. (2014) TTF-1 expression in breast carcinoma: an unusual but real phenomenon. *Histopathology* 64(4):504-11
164. Tsai LH, Chen PM, Cheng YW, et al. (2014) LKB1 loss by alteration of the NKX2-1/p53 pathway promotes tumor malignancy and predicts poor survival and relapse in lung adenocarcinomas. *Oncogene* 33(29):3851-60
165. Tan D, Li Q, Deeb G, et al. (2003) Thyroid transcription factor-1 expression prevalence and its clinical implications in non-small cell lung cancer: a high-throughput tissue microarray and immunohistochemistry study. *Hum Pathol* 34(6):597-604
166. Haque AK, Syed S, Lele SM, et al. (2002) Immunohistochemical study of thyroid transcription factor-1 and HER2/neu in non-small cell lung cancer: strong thyroid transcription factor-1 expression predicts better survival. *Appl Immunohistochem Mol Morphol* 10(2):103-9
167. Pelosi G, Frassetto F, Pasini F, et al. (2001) Immunoreactivity for thyroid transcription factor-1 in stage I non-small cell carcinomas of the lung. *Am J Surg Pathol* 25(3):363-72
168. Yamaguchi T, Hosono Y, Yanagisawa K, et al. (2013) NKX2-1/TTF-1: an enigmatic oncogene that functions as a double-edged sword for cancer cell survival and progression. *Cancer Cell* 23(6):718-23
169. Mu D (2013) The complexity of thyroid transcription factor 1 with both pro- and anti-oncogenic activities. *J Biol Chem* 288(35):24992-5000
170. Hamdan H, Liu H, Li C, et al. (1998) Structure of the human Nkx2.1 gene. *Biochim Biophys Acta* 1396(3):336-48
171. Kwei KA, Kim YH, Girard L, et al. (2008) Genomic profiling identifies TTF1 as a lineage-specific oncogene amplified in lung cancer. *Oncogene* 27(25):3635-40
172. Hirai H, Arai T, Okada M, et al. (2010) MK-1775, a small molecule Wee1 inhibitor, enhances anti-tumor efficacy of various DNA-damaging agents, including 5-fluorouracil. *Cancer Biol Ther* 9(7):514-22
173. Bridges KA, Hirai H, Buser CA, et al. (2011) MK-1775, a novel Wee1 kinase inhibitor, radiosensitizes p53-defective human tumor cells. *Clin Cancer Res* 17(17):5638-48
174. Rajeshkumar NV, De Oliveira E, Ottenhof N, et al. (2011) MK-1775, a potent Wee1 inhibitor, synergizes with gemcitabine to achieve tumor regressions, selectively in p53-deficient pancreatic cancer xenografts. *Clin Cancer Res* 17(9):2799-806
175. Osman AA, Monroe MM, Ortega Alves MV, et al. (2015) Wee-1 kinase inhibition overcomes cisplatin resistance associated with high-risk TP53 mutations in head and neck cancer through mitotic arrest followed by senescence. *Mol Cancer Ther* 14(2):608-19
176. Xu L, Huang CC, Huang W, et al. (2002) Systemic tumor-targeted gene delivery by anti-transferrin receptor scFv-immunoliposomes. *Mol Cancer Ther* 1(5):337-46
177. Xu L, Tang WH, Huang CC, et al. (2001) Systemic p53 gene therapy of cancer with immunoliposomes targeted by anti-transferrin receptor scFv. *Mol Med* 7(10):723-34
178. Camp ER, Wang C, Little EC, et al. (2013) Transferrin receptor targeting nanomedicine delivering wildtype p53 gene sensitizes pancreatic cancer to gemcitabine therapy. *Cancer Gene Ther* 20(4):222-8
179. Kim SS, Rait A, Kim E, et al. (2015) A tumor-targeting p53 nanodelivery system limits chemoresistance to temozolomide prolonging survival in a mouse model of glioblastoma multiforme. *Nanomedicine* 11(2):301-11
180. Pirolo KF, Nemunaitis J, Leung PK, et al. (2016) Safety and Efficacy in Advanced Solid Tumors of a Targeted Nanocomplex Carrying the p53 Gene Used in Combination with Docetaxel: A Phase 1b Study. *Mol Ther* 24(9):1697-706
181. Hajdenberg et al., 2012; ASCO Abstract e15010
182. Leijen S, van Geel RM, Pavlick AC, et al. (2016) Phase I Study Evaluating WEE1 Inhibitor AZD1775 As Monotherapy and in Combination With Gemcitabine, Cisplatin, or Carboplatin in Patients With Advanced Solid Tumors. *J Clin Oncol* ePub Sep 2016
183. Oza et al., 2015; ASCO Abstract 5506
184. Leijen et al., 2015; ASCO Abstract 2507
185. Ma CX, Cai S, Li S, et al. (2012) Targeting Chk1 in p53-deficient triple-negative breast cancer is therapeutically beneficial in human-in-mouse tumor models. *J Clin Invest* 122(4):1541-52
186. Mogi A, Kuwano H (2011) TP53 mutations in non-small cell lung cancer. *J Biomed Biotechnol* 2011:583929
187. Tekpli X, Landvik NE, Skaug V, et al. (2013) Functional effect of polymorphisms in 15q25 locus on CHRNA5 mRNA, bulky DNA adducts and TP53 mutations. *Int J Cancer* 132(8):1811-20
188. Vignot S, Frampton GM, Soria JC, et al. (2013) Next generation sequencing reveals high concordance of recurrent somatic alterations between primary tumor and metastases from patients with non-small cell lung cancer. *J Clin Oncol* 31(17): 2167-72
189. Maeng CH, Lee HY, Kim YW, et al. (2013) High-throughput molecular genotyping for small biopsy samples in advanced non-small cell lung cancer patients. *Anticancer Res* 33(11): 5127-33
190. Cortot AB, Younes M, Martel-Planche G, et al. (2014) Mutation of TP53 and alteration of p14(arf) expression in EGFR- and KRAS-mutated lung adenocarcinomas. *Clin Lung Cancer* 15(2): 124-30
191. Itakura M, Terashima Y, Shingyoji M, et al. (2013) High CC chemokine receptor 7 expression improves postoperative prognosis of lung adenocarcinoma patients. *Br J Cancer* 109(5):1100-8
192. Seo JS, Ju YS, Lee WC, et al. (2012) The transcriptional landscape and mutational profile of lung adenocarcinoma. *Genome Res* 22(11):2109-19
193. Brown CJ, Lain S, Verma CS, et al. (2009) Awakening guardian angels: drugging the p53 pathway. *Nat Rev Cancer* 9(12): 862-73
194. Joerger AC, Fersht AR (2008) Structural biology of the tumor suppressor p53. *Annu Rev Biochem* 77:557-82
195. Kato S, Han SY, Liu W, et al. (2003) Understanding the function-structure and function-mutation relationships of p53 tumor suppressor protein by high-resolution missense mutation analysis. *Proc Natl Acad Sci USA* 100(14):8424-9
196. Kamada R, Nomura T, Anderson CW, et al. (2011) Cancer-associated p53 tetramerization domain mutants: quantitative analysis reveals a low threshold for tumor suppressor inactivation. *J Biol Chem* 286(1):252-8
197. Bougeard G, Renaux-Petel M, Flaman JM, et al. (2015) Revisiting Li-Fraumeni Syndrome From TP53 Mutation Carriers. *J Clin Oncol* 33(21):2345-52
198. Sorrell AD, Espenschied CR, Culver JO, et al. (2013) Tumor protein p53 (TP53) testing and Li-Fraumeni syndrome: current status of clinical applications and future directions. *Mol Diagn Ther* 17(1):31-4
199. Nichols KE, Malkin D, Garber JE, et al. (2001) Germline p53 mutations predispose to a wide spectrum of early-onset cancers. *Cancer Epidemiol Biomarkers Prev* 10(2):83-7
200. Taubert H, Meye A, Würl P (1998) Soft tissue sarcomas and p53 mutations. *Mol Med* 4(6):365-72



**QRF#** XXXXXXXX

**APPENDIX**
**References**

201. Kleihues P, Schäuble B, zur Hausen A, et al. (1997) Tumors associated with p53 germline mutations: a synopsis of 91 families. *Am J Pathol* 150(1):1-13
202. Gonzalez KD, Noltner KA, Buzin CH, et al. (2009) Beyond Li Fraumeni Syndrome: clinical characteristics of families with p53 germline mutations. *J Clin Oncol* 27(8):1250-6
203. Lalloo F, Varley J, Ellis D, et al. (2003) Prediction of pathogenic mutations in patients with early-onset breast cancer by family history. *Lancet* 361(9363):1101-2
204. Ohe et al., 2015; ASCO Abstract 8061
205. Seto T, Kiura K, Nishio M, et al. (2013) CH5424802 (RO5424802) for patients with ALK-rearranged advanced non-small-cell lung cancer (AF-001JP study): a single-arm, open-label, phase 1-2 study. *Lancet Oncol* 14(7):590-8
206. Gadgeel SM, Gandhi L, Riely GJ, et al. (2014) Safety and activity of alectinib against systemic disease and brain metastases in patients with crizotinib-resistant ALK-rearranged non-small-cell lung cancer (AF-002JG): results from the dose-finding portion of a phase 1/2 study. *Lancet Oncol* 15(10):1119-28
207. Peters S, Camidge DR, Shaw AT, et al. (2017) Alectinib versus Crizotinib in Untreated ALK-Positive Non-Small-Cell Lung Cancer. *N Engl J Med* 377(9):829-838
208. Hida T, Nokihara H, Kondo M, et al. (2017) Alectinib versus crizotinib in patients with ALK-positive non-small-cell lung cancer (J-ALEX): an open-label, randomised phase 3 trial. *Lancet* 390(10089):29-39
209. Novello S, Mazières J, Oh JJ, et al. (2018) Alectinib versus chemotherapy in crizotinib-pretreated anaplastic lymphoma kinase (ALK)-positive non-small-cell lung cancer: results from the phase III ALUR study. *Ann Oncol* 29(6):1409-1416
210. Shaw AT, Gandhi L, Gadgeel S, et al. (2015) Alectinib in ALK-positive, crizotinib-resistant, non-small-cell lung cancer: a single-group, multicentre, phase 2 trial. *Lancet Oncol ePub Dec 2015*
211. Lin et al., 2018; ASCO Abstract 9093
212. Gainor JF, Sherman CA, Willoughby K, et al. (2015) Alectinib salvages CNS relapses in ALK-positive lung cancer patients previously treated with crizotinib and ceritinib. *J Thorac Oncol* 10(2):232-6
213. Ajimizu H, Kim YH, Mishima M (2015) Rapid response of brain metastases to alectinib in a patient with non-small-cell lung cancer resistant to crizotinib. *Med Oncol* 32(2):477
214. Ou SH, Sommers KR, Azada MC, et al. (2015) Alectinib induces a durable (>15 months) complete response in an ALK-positive non-small cell lung cancer patient who progressed on crizotinib with diffuse leptomeningeal carcinomatosis. *Oncologist* 20(2):224-6
215. Klemperner SJ, Ou SH (2015) Anaplastic lymphoma kinase inhibitors in brain metastases from ALK+ non-small cell lung cancer: hitting the target even in the CNS. *Chin Clin Oncol* 4(2):20
216. Dempke WC, Edvardsen K, Lu S, et al. (2015) Brain Metastases in NSCLC - are TKIs Changing the Treatment Strategy? *Anticancer Res* 35(11):5797-806
217. Lin JJ, Kennedy E, Sequist LV, et al. (2016) Clinical Activity of Alectinib in Advanced RET-Rearranged Non-Small-Cell Lung Cancer. *J Thorac Oncol ePub Aug 2016*
218. Kim et al., 2016; ASCO Abstract 9007
219. Camidge et al., 2016; World Conference on Lung Cancer P3.02a-013
220. Zhang S, Anjum R, Squillace R, et al. (2016) The Potent ALK Inhibitor Brigatinib (AP26113) Overcomes Mechanisms of Resistance to First- and Second- Generation ALK Inhibitors in Preclinical Models. *Clin Cancer Res*
221. Siaw JT, Wan H, Pfeifer K, et al. (2016) Brigatinib, an anaplastic lymphoma kinase inhibitor, abrogates activity and growth in ALK-positive neuroblastoma cells, Drosophila and mice. *Oncotarget* 7(20):29011-22
222. Kim DW, Tiseo M, Ahn MJ, et al. (2017) Brigatinib in Patients With Crizotinib-Refractory Anaplastic Lymphoma Kinase-Positive Non-Small-Cell Lung Cancer: A Randomized, Multicenter Phase II Trial. *J Clin Oncol* :JCO2016715904
223. Felip et al., 2015; ASCO Abstract 8060
224. Soria JC, Tan DS, Chiari R, et al. (2017) First-line ceritinib versus platinum-based chemotherapy in advanced ALK-rearranged non-small-cell lung cancer (ASCEND-4): a randomised, open-label, phase 3 study. *Lancet* 389(10072):917-929
225. Felip et al., 2015; ELCC Abstract 141PD
226. Scagliotti et al., 2016; ESMO Abstract LBA42\_PR
227. Crinò L, Ahn MJ, De Marinis F, et al. (2016) Multicenter Phase II Study of Whole-Body and Intracranial Activity With Ceritinib in Patients With ALK-Rearranged Non-Small-Cell Lung Cancer Previously Treated With Chemotherapy and Crizotinib: Results From ASCEND-2. *J Clin Oncol* 34(24):2866-73
228. Kodityal S, Elvin JA, Squillace R, et al. (2016) A novel acquired ALK F1245C mutation confers resistance to crizotinib in ALK-positive NSCLC but is sensitive to ceritinib. *Lung Cancer* 92:19-21
229. Katayama R, Friboulet L, Koike S, et al. (2014) Two novel ALK mutations mediate acquired resistance to the next-generation ALK inhibitor alectinib. *Clin Cancer Res* 20(22):5686-96
230. Ou SH, Greenbowe J, Khan ZU, et al. (2015) I1171 missense mutation (particularly I1171N) is a common resistance mutation in ALK-positive NSCLC patients who have progressive disease while on alectinib and is sensitive to ceritinib. *Lung Cancer* 88(2):231-4
231. Lu et al., 2016; ASCO Abstract 9058
232. Solomon BJ, Mok T, Kim DW, et al. (2014) First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *N Engl J Med* 371(23):2167-77
233. Butrynski JE, D'Adamo DR, Hornick JL, et al. (2010) Crizotinib in ALK-rearranged inflammatory myofibroblastic tumor. *N Engl J Med* 363(18):1727-33
234. Pérot G, Soubeyran I, Ribeiro A, et al. (2014) Identification of a recurrent STRN/ALK fusion in thyroid carcinomas. *PLoS ONE* 9(1):e87170
235. Blackhall F, Kim DW, Besse B, et al. (2014) Patient reported outcomes and quality of life in PROFILE 1007: a randomized trial of crizotinib compared with chemotherapy in previously treated patients with ALK-positive advanced non-small-cell lung cancer. *Thorac Oncol* 9(11):1625-33
236. Shaw et al., 2016; ASCO Abstract 9066
237. Solomon BJ, Cappuzzo F, Felip E, et al. (2016) Intracranial Efficacy of Crizotinib Versus Chemotherapy in Patients With Advanced ALK-Positive Non-Small-Cell Lung Cancer: Results From PROFILE 1014. *J Clin Oncol* 34(24):2858-65
238. Costa DB, Shaw AT, Ou SH, et al. (2015) Clinical Experience With Crizotinib in Patients With Advanced ALK-Rearranged Non-Small-Cell Lung Cancer and Brain Metastases. *J Clin Oncol* 33(17):1881-8
239. Johung KL, Yeh N, Desai NB, et al. (2016) Extended Survival and Prognostic Factors for Patients With ALK-Rearranged Non-Small-Cell Lung Cancer and Brain Metastasis. *J Clin Oncol* 34(2):123-9
240. Ou SH, Jänne PA, Bartlett CH, et al. (2014) Clinical benefit of continuing ALK inhibition with crizotinib beyond initial disease progression in patients with advanced ALK-positive NSCLC. *Ann Oncol* 25(2):415-22
241. Sledge GW, Toi M, Neven P, et al. (2017) MONARCH 2: Abemaciclib in Combination With Fulvestrant in Women With HR+/HER2- Advanced Breast Cancer Who Had Progressed While Receiving Endocrine Therapy. *J Clin Oncol* 35(25):2875-2884
242. Dickler MN, Tolane SM, Rugo HS, et al. (2017) MONARCH 1, A Phase II Study of Abemaciclib, a CDK4 and CDK6 Inhibitor, as a Single Agent, in Patients With Refractory HR(+)/HER2(-) Metastatic Breast Cancer. *Clin Cancer Res* 23(17):5218-5224
243. Kim et al., 2015; ASCO Abstract 8047
244. Dickson MA, Tap WD, Keohan ML, et al. (2013) Phase II trial of the CDK4 inhibitor PD0332991 in patients with advanced CDK4-amplified well-differentiated or dedifferentiated liposarcoma. *J Clin Oncol* 31(16):2024-8
245. Turner NC, Ro J, André F, et al. (2015) Palbociclib in Hormone-Receptor-Positive Advanced Breast Cancer. *N Engl J Med ePub Jun 2015*
246. DeMichele A, Clark A, Tan KS, et al. (2014) CDK 4/6 Inhibitor Palbociclib (PD0332991) in Rb+ Advanced Breast Cancer: Phase II Activity, Safety and Predictive Biomarker Assessment. *Clin Cancer Res ePub Dec 2014*
247. Gopalan et al., 2014; ASCO Abstract 8077
248. Goldman et al., 2014; ASCO Abstract 8026
249. Yamada et al., 2015; AACR-NCI-EORTC Abstract B31
250. Geoerger B, Bourdeaut F, DuBois SG, et al. (2017) A Phase I Study of the CDK4/6 Inhibitor Ribociclib (LEE011) in Pediatric Patients with Malignant Rhabdoid Tumors, Neuroblastoma, and Other Solid Tumors. *Clin Cancer Res*